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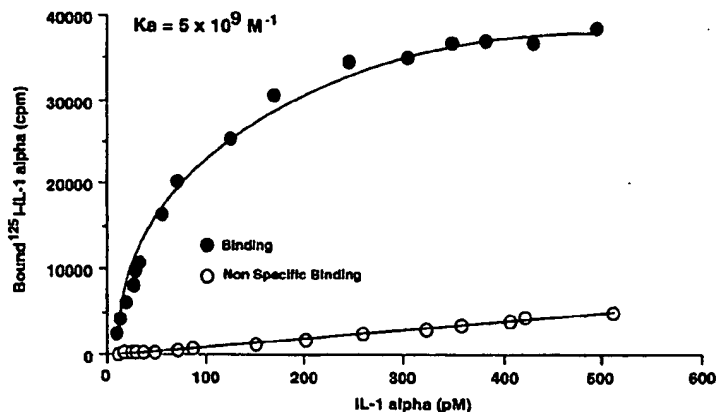
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(54) **Human monoclonal antibodies against human cytokines and methods of making and using such antibodies.**

(57) **Human monoclonal antibodies against a human cytokine (such as a human interleukin, e.g., human IL-1 α) and fragments of such antibodies are disclosed. Also disclosed are pharmaceutical compositions and methods employing the human monoclonal antibodies and fragments, methods for screening for human monoclonal antibodies against a human protein, methods for producing a cDNA library enriched in DNA encoding V_H and/or V_L chains of a human monoclonal antibody, cell lines for making the human monoclonal antibodies, and isolated DNA for making the human monoclonal antibodies and fragments of the invention.**

FIGURE 1



Exemplified
w/ Abs to
h TNF α

EP 0 659 766 A1

BACKGROUND OF THE INVENTION

The present invention relates to human monoclonal antibodies against human cytokines and methods of making, identifying and using such antibodies, preferably human monoclonal antibodies against human cytokines or lymphokines such as IL-1 α , IL-1 β , IL-4, IL-5, IL-6, IL-8, IL-10, TNF- α , etc.

The applicability of human monoclonal antibodies (HuMAbs), especially HuMAbs to human cytokines, in therapy holds great promise. See, for example, Griffiths et al., *EMBO J.*, 12 : 725-734 (1993) and the review in Larrick et al., *J. Biol. Response Modif.*, 5 : 379 (1986). However, the production of useful HuMAbs against human cytokines has proven difficult.

Specifically, while the possible existence in human serum of autoantibodies to human cytokines is mentioned in numerous articles (Suzuki et al., *J. Immunol.*, 145 : 2140-2146 (1990) (IL-1 α); Hansen et al., *Immunol. Letters*, 30 : 133-140 (1991) (IL-1 α); Bendtzen et al., *Immunol. Today*, 11 : 167-169 (1990) (IL-1 α and TNF- α); Bendtzen et al., *Immunol. Today*, 10 : 222 (1989) (IL-1 α and TNF- α); Saurat et al., *J. Allergy Clin. Immunol.*, 88 : 244-256 (1991) (IL-1 α); Suzuki et al., *Clin. Exp. Immunol.*, 85 : 407-412 (1991) (IL-1 α); Sunder-Plassmann et al., *Kidney International*, 40 : 787-791 (1991) (IL-1 α); Gallay et al., *Eur. Cytokine Netw.*, 2 : 329-338 (1991) (IL-1 α and IL-1 β); Mae et al., *Lymphokine Cryokine Res.*, 10 : 61-68 (1991) (IL-1 α); Fomsgaard et al., *Scand. J. Immunol.*, 30 : 219 (1989) (TNF- α); Hansen et al., *Scand. J. Immunol.*, 33 : 777-781 (1991) (IL-6); Crabtree et al., *Scand. J. Immunol.*, 37 : 65-70 (1993) (IL-8); Bost et al., *Immunology*, 65 : 611-615 (1988) (IL-2); Ross et al., *Clin. Exp. Immunol.*, 82 : 57-62 (1990) (IFN- α 2b and IFN- γ); and Caruso et al., *J. Immunol.*, 144 : 685-690 (1990) (IFN- γ)), no one has been able to produce an isolated and purified HuMAb to a human cytokine, especially a HuMAb having high affinity, e.g., a K_a of above about 10^9 M $^{-1}$. Some of the reasons are pointed out in the Griffiths et al. article:

"Human monoclonal antibodies (mAbs) have huge potential for therapy, but are difficult to make by immortalizing B-lymphocytes. Furthermore, it is especially difficult to generate human mAbs directed against human antigens (anti-self antibodies), for example antibodies against soluble TNF to block septic shock, against membrane bound carcinoembryonic antigen to image colorectal carcinoma or against lymphocyte antigens to destroy tumour in lymphoma. This difficulty results from immunological tolerance mechanisms that prevent the antigen-driven expansion of B-cell clones with self specificities. After antibody gene rearrangement, virgin B-cells may display antibodies with self-reactivity, but tolerance mechanisms can lead to their deletion or to their anergy. It has been suggested that cells may be anergized if the antigen is soluble, but deleted if the antigen is membrane bound. B-cell tolerance does not seem to occur when concentrations of soluble antigen are low (in contrast to T-cell tolerance) and B-cells with poor affinities for antigen are not tolerized, even at higher antigen concentrations. Such non-tolerized B-cells are not usually expanded because they lack T-cell help, although proliferation can be induced artificially by using polyclonal B-cell activators.

It is estimated that 10-30% of B-lymphocytes in normal, healthy individuals are engaged in making autoantibodies. However, the 'natural autoantibodies' produced do not lend themselves to therapeutic use as they are often IgM, low affinity and polyreactive." (Citations omitted.)

Although the Griffiths et al. article speaks of "human self-antibodies with high specificity," only single chain V_H and V_L fragments are actually disclosed. Moreover, there is no disclosure in the article that any of the heavy/light chain combinations mentioned therein are actually from one human antibody. Moreover, the human antibody fragments disclosed all have relatively low affinities, i.e., K_a s below 2×10^7 M $^{-1}$ and most below 10^7 M $^{-1}$.

SUMMARY OF THE INVENTION

The present invention is directed to human monoclonal antibodies against a human cytokine and to fragments of such antibodies having an affinity for the cytokine of 10^8 M $^{-1}$ or greater. The human monoclonal antibody (sometimes referred to herein as a HuMAb) or fragment preferably binds to a human lymphokine, more preferably to a human interleukin, e.g., human IL-1 α , IL-1 β , IL-4, IL-5, IL-6, IL-8, IL-10, IL-11, IL-12, IL-13, especially IL-1 α . The human monoclonal antibody or fragment of the invention preferably has an affinity (K_a) to the human cytokine of greater than 10^8 M $^{-1}$. The human monoclonal antibody or fragment preferably neutralizes the activity of the human cytokine. Human monoclonal antibodies of the IgG class are particularly preferred.

Another aspect of the invention involves a human monoclonal antibody or a fragment thereof comprising at least one CDR of an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 and/or of an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2; or one or more somatic variants of such sequences.

A preferred embodiment of the invention relates to a human monoclonal antibody or a human IL-1 α binding fragment comprising:

a V_H segment having an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 or by a CDR somatic variant thereof, and/or

5 a V_L segment having an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2 or by a CDR somatic variant thereof.

Preferably, the antibody comprises a V_H segment having an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 and/or a V_L segment having an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2. More preferably, the antibody comprises V_H and V_L segments having the amino acid sequences defined by amino acid residues 1-122 of SEQ ID NO. 1 and by amino acid residues 1-108 of SEQ ID NO. 2, respectively, or comprises a CDR somatic variant of one or both of said amino acid sequences. Particularly preferred is an antibody having V_H and V_L segments of the amino acid sequences defined by amino acid residues 1-122 of SEQ ID NO. 1 and amino acid residues 1-108 of SEQ ID NO. 2, respectively, e.g., an antibody of the human IgG₁ isotype.

15 Preferred fragments of the invention comprise a V_H segment having an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 and/or a V_L segment having an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2, e.g., a Fv, single chain Fv, Fab or F(ab')₂ fragment. Preferably, the IL-1 α binding fragment of the invention has an affinity of 10⁷ M⁻¹ or greater, more preferably of 10⁸ M⁻¹ or greater.

20 Another aspect of the invention involves isolated nucleic acids (DNAs) which encode a human monoclonal antibody or fragment in accordance with the present invention. Preferably, the isolated nucleic acid comprises:

a nucleotide sequence defined by base numbers 58-423 of SEQ ID NO. 1 or by a CDR encoding somatic variant thereof, or a functional equivalent of such a nucleotide sequence, and/or

25 a nucleotide sequence defined by base numbers 67-390 of SEQ ID NO. 2 or by a CDR encoding somatic variant thereof; or

a functional equivalent of one or both of said nucleotide sequences.

In a preferred embodiment, the isolated nucleic acid comprises a nucleotide sequence defined by base numbers 58-423 of SEQ ID NO. 1 and/or base numbers 67-390 of SEQ ID NO. 2.

30 Still other aspects of the invention relate to a pharmaceutical composition comprising at least one human monoclonal antibody or fragment in accordance with the invention and a pharmaceutically acceptable carrier, and to the use of an anti-IL-1 α HuMAb or IL-1 α binding fragment of the invention to treat inflammation.

The invention also includes a method for screening a solution for a desired human monoclonal antibody against a human protein comprising

(1) contacting the solution with labeled protein and polyclonal or monoclonal anti-human Ig (i.e., anti-IgA, IgD, IgE, IgG and/or IgM) coupled to a substrate or with labeled protein and protein G coupled to a substrate; and

40 (2) determining if a desired human monoclonal antibody is present in the solution by detecting labeled protein in any immunoprecipitated product.

Preferably, the solution is a collection of supernatants from a human B cell mixture. In a preferred embodiment, the solution is screened using either polyclonal or monoclonal anti-human Ig coupled to a substrate or with protein G coupled to a substrate. These screening methods can be used to prepare and identify a purified mixture of human B cells or a single human B cell clone by the steps of

45 serially diluting a human B cell mixture giving a positive result in the screen to provide a purified mixture of human B cells or single B cell clones;

culturing said purified mixture of human B cells or single human B cell clones; and

50 screening supernatants from said cultured purified mixture of human B cells or single B cell clones by the above methods to determine if the desired human monoclonal antibody is present in the supernatants of said cultured purified mixture of human B cells or single B cell clones.

Still another aspect of the invention involves a method of producing a cDNA library enriched in DNA encoding a V_H and/or V_L chain of a human monoclonal antibody against a desired antigen comprising the steps of:

55 producing a CD40-crosslinked and EBV-transformed, immortalized and/or activated B cell population containing immortalized and/or activated B cells expressing said human monoclonal antibody;

cloning subpopulations of said immortalized and/or activated B cell population and identifying a subpopulation which contains immortalized and/or activated B cells expressing said human monoclonal antibody;

preparing a cDNA library using the mRNA from said subpopulation to create a repertoire of DNAs encoding at least the V_H and/or V_L chain of the human monoclonal antibodies expressed by said subpopulation of immortalized and/or activated B cells.

Preferably, this method further comprises:

- 5 identifying DNA within said library that encodes at least the V_H and/or V_L chain of the desired human monoclonal antibody; and

using said DNA to produce a human monoclonal antibody against the desired antigen or an antigen-binding fragment of such an antibody.

- 10 In one such embodiment, a population or subpopulation which contains immortalized and/or activated B cells expressing said human monoclonal antibody is identified by the screening method described above.

- Preferably, the repertoire of DNAs are included in vectors capable of displaying said V_H and/or V_L chain on the surface of a host cell, host cells are transformed with said vectors, and host cells that display a V_H and/or V_L chain that binds to the desired antigen are identified by affinity binding to the desired antigen. The DNAs that encode said V_H and V_L chains that bind to the desired antigen can then be operatively linked
- 15 to DNA encoding any necessary constant region chains for a human immunoglobulin so as to create a DNA sequence encoding a heavy chain of a human monoclonal antibody against the desired antigen and a DNA sequence encoding a light chain of a human monoclonal antibody against the desired antigen.

- Other important aspects of the invention include (1) a human B cell line established by EBV-transformation and CD40-crosslinking, which established cell line (preferably an antibody-producing clone)
- 20 produces a human monoclonal antibody against a human cytokine, and (2) a process for making a human monoclonal antibody against a human cytokine comprising the steps of

- establishing an immortalized and/or activated human B cell population from a patient having antibodies that bind to the human cytokine, said immortalization and/or activation comprising infecting the B cells with Epstein-Barr virus and crosslinking the CD40 of such B cells;

- 25 culturing said immortalized and/or activated B cells;

isolating multiple clones from such immortalized and/or activated B cells, each of which clones secretes a human monoclonal antibody that binds to the cytokine; and

- using one or more of such clones to produce one or more human monoclonal antibody or a fragment thereof.

- 30 In this process nucleic acid encoding the human monoclonal antibody or fragment is preferably used to produce the desired antibody or fragment. Alternatively, the clone produced in the process is hybridized with a myeloma or heteromyeloma cell to produce a hybridoma that proliferates in culture and produces the desired antibody.

35 BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a graphical representation showing the amount of bound ^{125}I -IL-1 α (cpm) versus the concentration of IL-1 α (pM) in the "X3 Affinity for Human IL-1 α " assay described below.

- Figure 2 is a graphical representation showing the amount of ^{125}I -IL-1 α bound on EL4 cells (cpm)
- 40 versus the concentration of HuMAb X3 ($\mu\text{g/ml}$) in the "Inhibition of Human IL-1 α Receptor Binding" assay described below.

- Figure 3 is a graphical representation showing the amount of ^{125}I -IL-1 α precipitated (cpm) versus the concentration of HuMAb X3 ($\mu\text{g/ml}$) in the "Cross-Reactivity of the Human Monoclonal Antibody X3" assay described below using excess human IL-1 β , human IL-1Ra or human IL-1 α to protect against im-
- 45 munoprecipitation by HuMAb X3.

Figure 4A is a graphical representation showing the EL4/CTLL proliferation - ^3H TdR uptake (cpm) versus the concentration of IL-1 α (pg/ml) in the "Inhibition of Human IL-1 α -induced IL-2 Secretion by EL4 Cells" assay described below.

- Figure 4B is a graphical representation showing the EL4/CTLL proliferation (% of response) in the presence of human IL-1 β or human IL-1 α versus the concentration of HuMAb X3 ($\mu\text{g/ml}$) in the "Inhibition of Human IL-1 α -induced IL-2 Secretion by EL4 Cells" assay described below.

Figure 5A is a graphical representation showing the IL-6 production (ng/ml) in the presence of either 0.1 $\mu\text{g/ml}$ or 1 $\mu\text{g/ml}$ of HuMAb X3 versus the concentration of IL-1 α (pg/ml) in the "Inhibition of Human IL-1 α -induced IL-6 Production by Human Synoviocytes" assay described below.

- Figure 5B is a graphical representation showing the IL-6 production (% of response) in the presence of either human IL-1 β or IL-1 α versus the concentration of HuMAb X3 ($\mu\text{g/ml}$) in the "Inhibition of Human IL-1 α -induced IL-6 Production by Human Synoviocytes" assay described below.

Figures 6A and 6B are graphical representations showing the EL4/CTLL proliferation (cpm $\times 10^{-3}$) in the presence of rabbit anti-IL-1 β , rabbit anti-IL-1 α , or two concentrations of HuMAb X3 versus the amount of PFA-fixed Monocytes per well without and with LPS activation, respectively, in the "Inhibition of Membrane Associated Human IL-1 α Activity" assay described below.

- 5 Figures 7A, 7B, 7C and 7D are graphical representations showing the production of IL-6 (ng/ml) in the presence of CTL IgG4 \times , IL-1Ra or HuMAb X3 versus the amount of Monocytes/well, PFA-Monocytes/well, LPS-Monocytes/well and PFA/LPS-Monocytes/well, respectively, in the "Inhibition of IL-6 production in cocultures of synoviocytes and monocytes" assay described below.

- Figure 8A is a graphical representation showing the amount of 125 I-IL-1 α (cpm) precipitated in the "Standard Immunoprecipitation Protocol with Protein G" assay described below with various antibody materials, including the natural HuMAb X3 and the recombinant light and heavy chains from the HuMAb X3.

- Figure 8B is a graphical representation showing the amount of 125 I-IL-1 α (cpm) precipitated in the "Cross-Reactivity of the Human Monoclonal Antibody X3" assay described below using excess human IL-1 β , human IL-1Ra and human IL-1 α to protect against immunoprecipitation by natural HuMAb X3 or recombinant HuMAb X3.

DETAILED DESCRIPTION OF THE INVENTION

- The invention may employ a B cell population including resting B cells which retain their surface bound immunoglobulin and/or activated B cells which secrete HuMAbs. If desired, the B cell population may be sorted to select for activated B cells or for resting B cells, e.g., as described below and in WO 91/09115.

- A starting human B cell population for use in providing a human anti-cytokine HuMAb (or a subsequence thereof that binds to the cytokine) in accordance with the present invention can be identified by means conventional in the art, e.g., by the methods described in the articles listed in the Background of the Invention section above. A small amount of blood can be taken from patients and tested for Ig against the desired cytokine, e.g., by ELISA, radioimmunoprecipitation assay, western blotting, etc. Patients who react positively are sources of B cells that can be used to immortalize and isolate a clone producing the desired HuMAb as described further below. A larger sample for cloning can then be taken from each patient identified by the above procedures.

- Suitable sources of B cells from a selected patient include peripheral blood, tonsils, adenoid tissue, spleen (in the case of removal for another medical necessity) or any other source of B cells from the body. Typically, peripheral blood is employed as the B cell source.

- The blood is first treated to separate the peripheral blood lymphocytes (PBLs) from the red blood cells and platelets by means conventional in the art. For example, the peripheral blood may be diluted with an appropriate isotonic medium, e.g., RPMI 1640 medium (cat. 041-01870 M. Gibco, USA). The diluted blood is loaded onto a suitable separation medium such as FICOLL® (available from Pharmacia, Sweden). After centrifugation, the PBLs may be aspirated from the interface between the plasma and the FICOLL®. The purified PBLs may be frozen in liquid nitrogen for later use. The plasma is then analyzed by conventional techniques such as radioimmunoprecipitation assay, ELISA, western blotting, etc., to confirm the presence of significant amounts of the desired antibody (e.g., IgG $_1$, IgG $_2$, IgG $_3$, IgG $_4$, IgA, IgD, IgM and/or IgE antibody) against the cytokine of interest.

- The purified PBLs may be used directly or may be further enriched and/or sorted as discussed below. For example, T-cells may be removed by rosetting with 2-aminoethylisothiuronium bromide-treated sheep erythrocytes. Further selection for an antigen-specific B cell subpopulation can be carried out by a variety of techniques including panning, immunoadsorbent affinity chromatography, fluorescent-activated cell sorting (FACS), etc. These techniques are described for example in Casali et al., *Science*, **234** : 476-479 (1986); U.S. patent No. 4,325,706; and Mage, Hubbard et al., Parks et al. and Haegert, in *Meth. Enzymol.*, **108** : 118-124, 139-147, 197-241 and 386-392 (1984), respectively. The PBLs may also be treated with magnetic beads whose surface is coated with a material to selectively sort the desired B cells. Such beads may be coated, e.g., with anti-Ig isotype for the desired Ig to be separated, anti-surface antigen to select for non-naïve B cells, or a purified cytokine.

- The resulting enriched and/or sorted B cell population is then subjected to the B cell immortalization and/or activation process described in WO 91/09115. Briefly, the B cells are transformed with Epstein-Barr virus (EBV) and their CD40 molecules are crosslinked. Reference is made to WO 91/09115 for the variations that may be employed in this activation/immortalization process.

The treated B cell population may be washed by an appropriate isotonic medium (e.g., with RPMI 1640), pelleted and then resuspended in medium. The cells are then transformed with EBV by the addition of a suitable EBV strain, preferably a strain such as the one released by the B95.8 cell line available from

the ATCC (ATCC CRL 1612). The amount of EBV used may vary depending on the strain of the virus and the number of B cells to be transformed. For example, with a sample containing 14×10^6 non-sorted PBLs, 200 μ l of a concentrated EBV (strain B95.8) suspension is typically used. Incubation with the virus is typically carried out for about 1 to 24 hours, preferably for about 2 hours, at 37°C ; but other conditions may be employed, if desired.

The EBV-infected cells are preferably washed and resuspended in an appropriate enriched medium such as Yssel's modified Iscove's medium 15% Fetal Calf Serum (FCS) [Yssel, et al., *J. Immunol. Methods*, 72 : 219-227 (1984)]. The concentration of the PBLs in the suspension may vary depending, for example, on whether a sorting step for antigen-specific B cells was performed as described above. Lower concentrations can be employed when PBLs have been enriched in the desired B cells. Typical concentrations for non-selected PBLs are from about 1×10^3 to about 5×10^4 cells/ml. If the B cell population is first sorted as discussed above, the concentration may be decreased depending upon the efficiency of the sorting, e.g., up to about 1×10^2 cells/ml.

An agent capable of crosslinking CD40 antigen is added to the suspended cells. The crosslinking agent may include T-cells, other transfected cells expressing CD40 ligand or membranes therefrom. Other suitable agents are described in WO 91/09115. Preferably, the agent is an immobilized monoclonal antibody specific for the CD40 antigen, e.g., immobilized on irradiated fibroblasts expressing the human or murine Fc-gamma receptor (ATCC CRL 10680).

The monoclonal antibody to CD40 can be any which binds to the CD40 marker on the B cells of the suspension and also to the Fc-gamma receptor of the L-cells. Preferably, the monoclonal antibody is selected from MAb 89 and G28-5. These antibodies are described in Valle et al., *Eur. J. Immunol*, 19 : 1463-1467 (1989) and Ledbetter et al., *J. Immunol*, 138 : 788-794 (1987), respectively. The hybridoma corresponding to MAb 89 has been deposited with the European Collection of Animal Cell Cultures, PHLS Centre for Applied Microbiology and Research, Porton Down, Salisbury Wilts. SP4 OJG, U.K. under accession No. 89091401.

Typically, the PBLs, the CD40 antibody and the L-cells are simply mixed together in appropriate amounts. The CD40 antibody may be present in a concentration of from about 0.01 μ g/ml to about 50 μ g/ml, preferably from about 0.1 μ g/ml to about 5 μ g/ml, more preferably about 0.5 μ g/ml.

The treated cell suspension is divided among an appropriate number of wells of a tissue microplate to provide a suitable cell concentration for amplification and screening. If enriched suspensions are employed as the result of an antigen-selective screening as discussed above, fewer cells per well may be used.

Typically, the initial culture phase takes 10-20 days in the case of non-selected PBLs and 5 days or even less in the case of an antigen-specific enriched B cell population, which would allow an earlier detection of specific antibodies. During this phase, fresh medium is added as necessary. The duration of this initial culture phase is adjusted to allow detection of the antigen-specific B cells, while preventing them from being overgrown by non-specific B cells. A sample of supernatant from each well is screened by an appropriate assay for the desired HuMAb positive characteristics, e.g., by radioimmunoprecipitation assay, ELISA, western blotting, etc.

In a preferred screening method, supernatants are contacted with a labeled protein (e.g., radiolabeled with ^{125}I) and polyclonal or monoclonal anti-human Ig coupled to a substrate or insoluble support. The anti-human Ig can be a mixture of isotypes (i.e., IgG₁, IgG₂, IgG₃, IgG₄, IgA, IgD, IgE and/or IgM) or an individual isotype (e.g., IgG₄). Alternatively, if one is screening for an IgG HuMAb, the supernatants can be contacted with a labeled protein and protein G coupled to a substrate or insoluble support. The presence of the desired HuMAb is determined by detecting labeled protein in the immunoprecipitated product. The immunoprecipitation screens (with anti-human Ig and with protein G) may be employed serially.

Cell lines which test positive for the desired HuMAb characteristics are cloned (3-10 cells/well) and subcloned (0.5-1.0 cells/well) by techniques well-known in the art, e.g., by culturing in limiting dilution conditions for 7-20 days in additional medium as needed. Supernatants of the clones are screened by the procedures described above.

Positive clones are expanded in a larger volume and amplified by conventional incubation.

HuMAb can be purified from the supernatant of the amplified clones by conventional immunoglobulin purification methodology. For example, the HuMAb may be precipitated with solid ammonium sulfate, reconstituted in sterile water, and dialyzed extensively against a buffer such as phosphate buffer saline (PBS). The dialysate may then be applied to an immunoaffinity column, e.g., a column having anti-human Ig or Protein G covalently coupled to Sepharose. After washing, the desired HuMAb may be eluted from the column by any appropriate eluent, e.g., acidic buffer, chaotropic agents, etc. [for example, see in Current Protocols in Immunology, edited by John E. Coligan et al., John Wiley and Sons, New York].

By the term "human monoclonal antibody" as used herein, we mean to include HuMAbs that are isolated from human B cells as discussed above (e.g., whether the antibody is prepared by culturing the immortalized and/or activated human B cells or recombinantly from human B cell cDNAs encoding such a HuMAb and whether or not the antibody is bound to a molecule which can alter its biological activity, e.g., a receptor or ligand, an enzyme, a toxin, a carrier, etc.) and antibodies that are made by recombining the variable portions of a HuMAb of the present invention of one isotype (e.g., an IgG₄) with the constant region of a human antibody of another isotype (e.g., a human IgG₁, IgG₂, IgG₃, IgG₄, IgA, IgD, IgM or IgE). Recombinant methods for making these HuMAbs are described below.

By the terms "fragment" or "subsequence" of a HuMAb of the present invention, we mean an antibody fragment such as an Fab, F(ab')₂, Fv, single chain binding protein, or any other binding polypeptide which contains one or more complementarity determining regions (CDRs) of the variable region of a light or heavy chain of a HuMAb of the present invention (e.g., an Fab, Fv, CDR, etc. of a HuMAb in accordance with the present invention either alone or linked to any desired molecule which can alter its biological activity, e.g., a receptor or ligand, an enzyme, a toxin, a carrier, etc.). These fragments can be prepared by well-known methods. For example, fragments can be made from the full length HuMAb protein, e.g., by papain or pepsin cleavage, or by chemical oxidation, followed by separation of the resulting fragments. Alternatively, recombinant DNA technology may be used. For example, cDNA encoding the variable regions of both heavy and light chains may be engineered to produce the Fv portion of the HuMAb of the invention. See, for example, the methodology of U.S. patent No. 4,642,334 which may be employed.

By the terms "CDR somatic variant" and "CDR encoding somatic variant" as used herein we mean an amino acid or nucleic acid sequence corresponding to SEQ ID NO. 1 and/or SEQ ID NO. 2 or a subsequence of SEQ ID NO. 1 and/or SEQ ID NO. 2 containing at least one CDR or CDR encoding region thereof, but having at least one mutation, addition and/or deletion in one or more of the CDRs or CDR encoding region of the sequence or subsequence, such that an anti-IL-1 α human monoclonal antibody including said at least one mutation, addition and/or deletion has an IL-1 α binding affinity of 10⁸ M⁻¹ or greater, preferably 10⁹ M⁻¹ or greater.

By the term "affinity" as used herein, we mean the measure of the binding strength between an antigenic determinant and an antigen binding site of a human monoclonal antibody of the invention or a fragment thereof as measured by the affinity constant (K_a), e.g., by the method described below.

By the term "V_H segment" as used herein, we mean the variable region of the heavy chain of a human monoclonal antibody of the invention.

By the term "V_L segment" as used herein, we mean the variable region of the light chain of a human monoclonal antibody of the invention.

By the term "Fv fragment" as used herein, we mean an antigen binding fragment of an antibody that contains the variable regions of the heavy (V_H) and light (V_L) chains. Those V_H and V_L regions can be linked to form a single chain Fv (scFv).

By the term "Fab fragment" as used herein, we mean the antigen binding fragment resulting from the digestion with papain of a human monoclonal antibody of the invention.

By the term "F(ab')₂ fragment" as used herein, we mean the antigen binding fragment resulting from the digestion with pepsin of a human monoclonal antibody of the invention.

By the term "functional equivalent" as used herein, we mean a nucleic acid sequence that encodes the same amino acid sequence as the identified nucleic acid sequence.

By the term "activated" B cell and "activation" as used herein we mean a human B cell that has been CD40 crosslinked and EBV-transformed and expresses and secretes human antibodies.

The B cell clones of the invention may be used in conventional DNA recombinant methods to produce the HuMAbs of the invention or fragments thereof. For example, RNA from the B cell clones may be isolated according to the single-step method described by (Chomczynski, et al., *Anal. Biochem.*, 162 : 156-9 (1987). Briefly, about 10⁷ cells are lysed in guanidinium thiocyanate denaturing solution. After acidification of the mixture with 2M sodium acetate, pH4, RNA is extracted with phenol and chloroform/isoamyl alcohol. RNA is then precipitated with isopropanol, the RNA pellet is redissolved in denaturing solution, reprecipitated with isopropanol, and washed with 75% ethanol.

cDNA is obtained by reverse transcription, e.g., using the Superscript Reverse Transcriptase Kit (cat. 20898 BRL, Gaithersburg, MD, USA), with oligo dT₁₂₋₁₈ primers (Cat. 27.7858-01, Pharmacia, Upsalla, Sweden). The cDNA is then used as template in a polymerase chain reaction (PCR). The primers may be designed to include restriction sites, to allow for the directional cloning of the PCR products. For the heavy chain, primers specific for the leader sequence of all the different human V_H families are used individually in conjunction with primers located at the 3' end of the constant region corresponding to the isotype previously determined by isotyping the HuMAb by ELISA or other appropriate method (e.g., radioim-

munoprecipitation assay, etc.). The light chain is amplified with individual combinations of primers corresponding to the 3' end of the kappa or lambda chain in conjunction with a series of primers annealing to the leader sequence of the V kappa or the V lambda genes. Thus, full length heavy and light chains starting at the initiation codon in the leader sequence and ending at the stop codon may be generated.

After appropriate restriction cleavage, both full length heavy and light chains can then be cloned in any appropriate expression vector designed to be compatible with the restricted PCR products. Appropriate vectors include for example baculovirus vectors and plasmids compatible with CHO cells or other host cells. Examples of suitable vectors and hosts are described in the review "Engineered antibody molecules" in *Immunol. Reviews*, 130 (1992). Heavy and light chains can be cloned individually in distinct vectors, or in tandem in one vector. The recombinant plasmids or viral vectors may be cloned in bacteria, and a few clones may be sequenced on both strands to check for the absence of alteration of the insert. One clone each for the heavy and the light chain, or one clone containing both chains may then be selected for expression in the appropriate host cells. Depending on the vector used, it will be introduced in appropriate prokaryotic or eukaryotic cells either by transfection or infection. After cloning the cells expressing the recombinant HuMAb, supernatant fluid from the cultured cells is collected, and the HuMAb therein can be purified, e.g. by immunoaffinity, HPLC or any other appropriate methods.

The full length PCR product for the heavy chain can be modified for example to replace the original heavy chain constant region by another one, so substituting, e.g., a human IgG₄ isotype to a human IgG₁, IgG₂, IgG₃, IgA, IgD, IgM or IgE isotype. This can be accomplished through a second PCR using as 5' primer the V_H leader specific primer, and as 3' primer a primer annealing to the 3' end of the V_H region (e.g., IgG₄) and including a tail corresponding to the 5' end of the desired constant region (e.g., IgG₁, IgG₂, etc.) cloned in an appropriate plasmid. After amplification, the heavy chain coding DNA generated may be digested with appropriate enzymes and ligated into the new expression vector (containing the sequence of the desired heavy chain). This will allow the production of a recombinant HuMAb with the variable region of the originally isolated HuMAb (e.g., the variable region from an IgG₄ HuMAb) and the constant region of a different human isotype (e.g., the constant region of a human IgG₁, IgG₂, etc.). This type of recombinant HuMAb will have the characteristic binding of the IgG₄ HuMAb, but will be able to display the effector functions normally associated with the human IgG₁, IgG₂, etc. isotype.

Hybridomas may also be made with the B cells of the invention by techniques conventional in the art. For example, the B cells of the invention may be fused with an appropriate myeloma cell or with a heterohybridoma cell to increase or stabilize the immunoglobulin secretion. See for example Kudo et al., *J. Immunol. Methods*, 145 : 119-125 (1991); Zanella et al., *J. Immunol. Methods*, 156 : 205-215 (1992); and Darveau et al., *J. Immunol. Methods*, 159 : 139-143 (1993).

The HuMAbs and fragments of the present invention may be used therapeutically to treat existing symptoms associated with the antigen of interest. For example, IL-1 α , IL-1 β and TNF- α are identified as inflammatory cytokines and thus HuMAbs to such cytokines or fragments of such HuMAbs may be useful to treat inflammation including chronic or acute inflammatory reactions such as rheumatoid arthritis, osteoarthritis, inflammation associated with asthma, inflammatory bowel disease, regulating fever associated with inflammation, pain relief in inflammation, etc. For example, bowel diseases may benefit from a treatment with an anti-IL-1 α HuMAb of the invention, e.g., HuMAb X3, as anti-IL-1 α antibodies abolished the crypt hyperplasia in the jejunum of mice suffering from graft-versus-host disease enteropathy, Mowat et al., *Immunology*, 80 : 110-115 (1993). Also, since IL-1 α has been suspected to play a role in psoriasis, Romero et al., *J. Invest. Dermatol.*, 93 : 518-522 (1989), an anti-IL-1 α HuMAb of the invention may be useful in this respect. Allergy may be another target for an anti-IL-1 α HuMAb of the invention both at the regulatory and effector levels. At a regulatory level, IL-1 has been shown to be involved in the differentiation of naive T lymphocytes into TH₂ T cells. At the effector level, clinical trials with soluble IL-1 receptor have shown a striking inhibition of wheal and flare reaction in allergen challenged allergic individuals. Furthermore, the anti IL-1 HuMAbs and anti IL-1-binding fragments of the present invention are antagonists to IL-1 and therefore will be useful for the same indication as other known IL-1 antagonists, which either are in clinical trials for or have been shown in the literature to be useful in models of septic (endotoxin) shock, experimental autoimmune encephalomyelitis, cerebral malaria, graft-versus-host disease and chronic myelogenous leukemia (see for review Dinarello, *Immunol. Reviews*, 127 : 119-146 (1992); and Dinarello et al., *N. Engl. J. Med.*, 328 : 106-113 (1993). Finally, IL-1 α has been reported to act as an autocrine growth stimulator for human thyroid and gastric carcinoma cells, Ito et al., *Cancer Res.*, 53 : 4102-4106 (1993), as well as adult T cell leukemias, Shirakawa et al., *Cancer Res.*, 49 : 1143-1147 (1989). Thus, an anti-IL-1 α HuMAb of the invention may be useful in the treatment of tumors.

Further examples of the utilities of other antagonists of and/or antibodies to other cytokines are described in Henderson et al., *TIPS*, 13 : 145-152 (1992) and in Mire-Sluis, *TIBTECH*, 11 : 74-77 (1993).

The HuMAbs and fragments of the present invention may also be used prophylactically to prevent or inhibit the occurrence of such symptoms associated with the antigen of interest.

The HuMAbs of the invention may be particularly useful, e.g., in treating chronic diseases, in view of the long half lives of HuMAbs (e.g., about 21 days Adair, *Immunol. Reviews*, 130 : 5-40 (1992)) compared to other cytokine antagonist (about 30 minutes for IL-1 receptor antagonist, Granowitz et al., *Cytokine*, 4 : 353-360 (1992)). This longer half life may allow a bimonthly or monthly administration of the HuMAb.

The HuMAb or fragment thereof of the present invention may be used alone or in combination with at least one other HuMAb or fragment to form a cocktail or with another antiinflammatory drug. For example, such a cocktail may include two or more of the HuMAbs of the invention, each of which binds to one or more epitopes on a cytokine of interest. When such a cocktail is employed, the proportions of the various HuMAbs or fragments may vary depending, for example, upon their binding characteristics.

The HuMAbs and fragments of the present invention are preferably administered in the form of a pharmaceutical composition containing a therapeutically or prophylactically effective amount of at least one such HuMAb or fragment in combination with a pharmaceutically acceptable carrier. Any appropriate carrier may be employed, i.e., a compatible, non-toxic material suitable for delivery of the HuMAb or fragment in the desired dosage form, e.g., oral, parenteral (subcutaneous, intramuscular or intravenous), or topical dosage forms. Suitable carriers include sterile water, sterile buffered water, sterile saline, etc. Special pharmaceutical compositions to insure a sustained release of the HuMAb and/or fragment may also be employed.

The concentration of the HuMAb or fragment of the invention in the pharmaceutical compositions may vary, e.g., from about 0.1 µg/ml to about 1 mg/ml, preferably from about 1 µg/ml to about 100 µg/ml. The concentration used will depend upon the number of HuMAbs and/or fragments thereof employed in the composition, their binding characteristics and the dosage form selected. The dose will be adjusted in a conventional manner by the skilled artisan to levels appropriate to achieve the desired result *in vivo*.

As mentioned above, the HuMAbs and/or fragments of the invention can be used therapeutically or prophylactically. Thus, the agent may be administered before the onset of symptoms or after the symptoms have appeared. The HuMAbs and/or fragments of the invention will be administered in a dose effective to provide the desired alleviation of symptoms. Amounts effective for this purpose will depend upon many factors, e.g., the severity of the symptoms in the patient.

The HuMAb or fragment of the invention may be administered in dosages of from about 0.001 µg/kg to about 1 mg/kg, e.g., about 0.01 µg/kg to about 1 µg/kg preferably, from about 0.01 µg/kg to about 0.1 µg/kg. The proper dosage of a HuMAb or fragment of the invention for a particular situation will be determined by using common practices in the art. Generally, treatment may be initiated with smaller dosages that are less than the optimum dose of the agent. Thereafter, the dosage may be increased by small increments until the optimum effect under the circumstances is reached. The amount and frequency of administration of the HuMAb or fragment of the invention will be regulated according to the judgment of the attending clinician considering such factors as age, condition and size of the patient as well as severity of the symptom being treated.

The HuMAbs and fragments of the invention may also be used for diagnostic purposes in the same manner as other antibodies and fragments are currently used in the art. For example, the HuMAbs and fragments of the invention can be used in assays for the cytokine to which they bind or in an immunopurification procedure to isolate an antigen to which it binds. The HuMAbs and fragments may be used labeled (e.g., with a radioisotope, fluorescent group, enzyme or other appropriate ligand) or unlabeled as is conventional in the art for the particular assay of interest (e.g., in a sandwich assay with a second labeled antibody). The HuMAbs and fragments may be used in agglutination assays, enzyme immunoassays, etc. They could for example be used to calibrate a dosage of cytokine-specific IgG in the serum or in any other biological fluid. Thus, the labeled or unlabeled forms of the HuMAbs and fragments of the invention may be employed as elements of kits for purposes of performing the desired assay.

The invention disclosed herein is exemplified by the following examples, which should not be construed to limit the scope of the disclosure. Alternative methods within the scope of the invention will be apparent to those skilled in the art.

Unless otherwise indicated, percentages for solids in solid mixtures, liquids in liquids, and solids in liquids are expressed on a weight/weight, volume/volume and weight/volume basis, respectively.

EXAMPLES**General Methods and Reagents**

5 Recombinant human IL-1 alpha (IL-1 α) and recombinant human IL-1 Receptor antagonist (IL-1Ra) were expressed in *E. coli* by standard methods and purified by ion-exchange and gel chromatographies.

Recombinant human IL-1 beta (IL-1 β) was purchased from Genzyme (Boston, MA).

¹²⁵I-labeled recombinant human IL-1 α (specific activity: 1200-2200 Ci/mmol) was from Du Pont De Nemours (Wilmington, DE).

10 Mouse monoclonal antibody to human IL-1 α and rabbit neutralizing antibodies against human IL-1 α and human IL-1 β were from Genzyme (Cambridge, MA).

Protein G (from Group C *Streptococcus* sp.) coupled to Sepharose 4B Φ , and anti-human Ig polyvalent immunoserum (IgG fraction) coupled to agarose were obtained from Sigma Chemical Co. (St Louis, MO).

15 Tissue culture media, fetal calf serum (FCS), L-glutamine, Hepes buffer, Phosphate Buffered Saline (PBS) were from GIBCO (Paisley, UK). Bovine serum albumin (BSA) was from Sigma Chemical Co. and gentamycin from Schering-Plough (Levallois-Perret, France).

The murine thymoma cell line EL4 (ATCC, TIB 181) was maintained in RPMI 1640 supplemented with 10% FCS, 2mM L-glutamine, 50 μ g/ml gentamycin and 5×10^{-5} M 2-mercaptoethanol (2-ME) in a humidified 37°C chamber with 5% CO₂.

20 The murine IL-2-dependent cytotoxic T cell line (CTLL-2) (ATCC, TIB 214) was maintained in RPMI 1640 supplemented with 10% FCS, 2mM L-glutamine, 50 μ g/ml gentamycin, 5×10^{-5} M 2-ME and 20 U/ml recombinant human IL-2 in a humidified 37°C chamber with 5% CO₂.

Human synoviocytes were isolated from rheumatoid synovial biopsies obtained from rheumatoid arthritis patients undergoing knee or wrist synovectomy, or joint replacement as described in Dechanet et al., *J. Immunol.*, 151 : 4908-4917 (1993). Fat and fibrous tissues were removed. The resulting fragments of synovium were finely minced into small pieces and digested with 4 mg/ml collagenase (Worthington, Freehold, NJ) in PBS for 2-3 hours at 37°C. After centrifugation, cells were resuspended in α -MEM (Gibco) medium supplemented with 2 mM L-glutamine, 20 mM Hepes buffer, 10% FCS and 50 μ g/ml gentamycin. Cells were cultured in 100 mm culture Petri dishes, in humidified 5% CO₂ atmosphere. After adherence for 30 48 hours, non-adherent cells were removed and adherent cells were cultured until confluence. Then cells were cultured in 150 cm² culture flasks after trypsin treatment. Synoviocytes were used between passage 3 to 8. They were negative for the expression of CD1, CD2, CD3, CD19, CD14 and HLA-DR as determined by flow cytometry analysis on a FACScan (Becton Dickinson, Sunnyvale, CA) after staining with specific fluorescein-isothiocyanate (FITC) conjugated monoclonal antibodies (mAbs) (Becton Dickinson, Mountain 35 View, CA).

The transformant Epstein-Barr virus (EBV), strain B 95.8, was produced by culturing transformed marmoset leukocytes (ATCC, CRL 1612) essentially as described by Miller and Lipman [*Proc. Natl. Acad. Sci. USA*, 70 : 190 (1973)].

40 The Itk⁻ transfected mouse fibroblastic L cell line (ATCC CRL 10680) stably expressing the human Fc γ receptor II (CDw32), and the mouse anti-human CD40 monoclonal antibody, mAb 89, were obtained as described in WO 91/09115.

Standard Immunoprecipitation Protocol with Protein G

45 In order to identify the presence of human antibodies (IgG) against human IL-1 α in biological samples (sera, plasma, etc.) or culture supernatants an immunoprecipitation assay was carried out using radio-labeled recombinant human IL-1 α and protein G-Sepharose as precipitating reagent. This assay allowed the identification of the four subclasses of human IgG (IgG₁, IgG₂, IgG₃ and IgG₄). Typically, 50 μ l of sera/plasma from patients or 50 μ l of culture supernatants (both used at appropriate dilution in PBS, 1% 50 BSA) were incubated for 45 minutes at room temperature with 50 μ l (50 pM) of human ¹²⁵I-IL-1 α (diluted in PBS 1% BSA) in a well of a 96-well filtration microplate MultiScreen-HA Φ (Millipore Co., Bedford, MA) which bottom was composed of a nitrocellulose membrane (HATF 0.45 μ m). Each sample was tested in duplicate. Then, 50 μ l of a dilution of protein G coupled to Sepharose 4B Φ (Sigma Chemical Co) (15 ml beads diluted to 50 ml in PBS 1% BSA) were added to each well and incubated for 45 minutes at room 55 temperature. The wells were then washed three times with PBS using a vacuum manifold (Millipore Co.) and the dried membranes were collected into appropriate vials using a special collector system (Millipore Co.). The radioactivity corresponding to the complexes ¹²⁵I-IL-1 α /anti-IL-1 α was counted in a Wizard gamma-counter (Wallac Oy, Turku, Finland). Positive and negative controls were performed for each plate

respectively using a rabbit anti-human IL-1 α antiserum (Genzyme, Cambridge, MA) or an unrelated antiserum. Specificity of the human 125 I-IL-1 α precipitation obtained with the tested samples was further confirmed through its inhibition by preincubation of these samples with a 100-fold excess of unlabeled recombinant human IL-1 α .

Derived Immunoprecipitation Protocols

The above standard immunoprecipitation assay has been modified in order to identify human antibodies to human IL-1 α of other isotypes than IgG or to better identify the IgG subclass and light chain of such antibodies contained in patient biological fluids, e.g., sera or culture supernatants. The principle of the different assays was the same as in the standard protocol, but the precipitating reagent, e.g. protein G-Sepharose, was changed. The following reagents were used: agarose beads coupled with goat polyclonal antibodies to human IgM, IgG and IgA (Sigma Chemical Co.); Affi-Gel 10 Φ gel (Bio-Rad laboratories, Richmond, CA) coupled, according to the manufacturer's instructions, with specific goat antibodies to human IgA heavy chain, human lambda light chain or human kappa light chain (Sigma Chemical Co.) or coupled with mouse monoclonal antibodies to human IgG $_1$, human IgG $_2$, human IgG $_3$ or human IgG $_4$ heavy chain (Calbiochem Co., La Jolla, CA).

These protocols may be employed with other antigens of interest by substituting an appropriately labeled antigen for the 125 I-IL-1 α in the assays.

Detection of Human Antibodies to Human IL-1 α in Human Biological Fluids

Detection of naturally occurring autoantibodies to human IL-1 α in biological fluids, e.g., sera or plasma, was performed by using the radioimmuno-precipitation assay described above. Blood samples from healthy donors or sick individuals (particularly patients suffering from autoimmune diseases, infectious diseases or neurologic disorders) were screened. Ten % of the samples from healthy donors (101/1009) contained IgG antibodies to human IL-1 α , as, when diluted 1:10, they significantly precipitated 125 I-labeled human recombinant IL-1 α with protein G-Sepharose. Some samples also contained IgA autoantibodies to human IL-1 α , as determined by immunoprecipitation of 125 I-labeled IL-1 α with appropriate anti-human IgA reagents coupled to beads. An increased frequency (15.9%, 59/370) of IgG anti-IL-1 α autoantibodies was observed in sera of patients with autoimmune diseases. Precipitation of radio-labeled human IL-1 α was specific since it was completely inhibited by pre-incubation of the positive samples with a 100-fold excess of unlabeled human IL-1 α .

Autoantibodies to human IL-1 α were titrated using the immunoprecipitation assay for serial dilutions of positive sera or plasma. Then, sera or plasma samples with high titres of anti-IL-1 α antibodies were also tested for their ability to inhibit the binding of human 125 I-IL-1 α to its receptors expressed on the murine thymoma EL4 cells. Serial dilutions (in RPMI 1640, 1% BSA, 20 mM Hepes) of positive sera or plasma were pre-incubated for 1 hour at 4°C with a fixed concentration (70 pM) of human 125 I-IL-1 α , in a final volume of 100 μ l. Experiments were performed in conical 1 ml Eppendorf tubes or in V-bottomed microtiter plates (Nunc, Roskilde, Denmark), and each sample was tested in triplicate. Then, 1×10^6 (100 μ l) of EL4 cells in RPMI 1640, 1% BSA, 20 mM Hepes were added to each tested point and incubated for 3 hours at 4°C. Cells were then washed and centrifuged three times at 4°C, and the radioactivity corresponding to cell-bound 125 I-IL-1 α was counted in a Wizard gamma-counter (Wallac). Non-specific binding was measured in the presence of a 100-fold excess of unlabeled human IL-1 α . Most of the sera or plasma containing autoantibodies to human IL-1 α were found to block human IL-1 α binding to its receptor. Patients were selected according to these criteria. Examples of the reactivity of 5 different sera are shown in Table 1 below.

Table 1

| Detection of IgG autoantibodies to human IL-1 α in human sera | | | |
|--|----------|--|---|
| Sera | Dilution | ¹²⁵ I-IL-1 α precipitated with Protein G (cpm) | ¹²⁵ I-IL-1 α bound to EL4 cells cpm (% inhibition) |
| none | | 787 | 4516 |
| negative | 1/5 | 1082 | 4490 |
| C | 1/5 | 3860 | 903 (80) |
| | 1/10 | 3844 | 2365 (48) |
| | 1/20 | 2554 | 3233 (28) |
| S | 1/5 | 3697 | 1685 (63) |
| | 1/10 | 2763 | 3156 (30) |
| | 1/20 | 2061 | 3864 (14) |
| T | 1/5 | 3392 | 2332 (48) |
| | 1/10 | 2692 | 2945 (35) |
| | 1/20 | 1953 | 3559 (21) |
| V | 1/5 | 4130 | 642 (86) |
| | 1/10 | 3741 | 655 (85) |
| | 1/20 | 2486 | 1976 (56) |
| X | 1/5 | 3866 | 701 (84) |
| | 1/10 | 3723 | 2122 (53) |
| | 1/20 | 2593 | 2666 (41) |

In a similar manner, autoantibodies against other cytokines in human biological fluids, such as sera, plasma, etc., are detected by substituting, for example, ¹²⁵I-TNF- α , ¹²⁵I-IL-1 β , ¹²⁵I-IL-6 or ¹²⁵I-IL-10 for ¹²⁵I-IL-1 α in the above procedure. Samples from such patients are then EBV-transformed, CD40 activated, screened and amplified by the methods described below to produce subpopulations and/or single clones of B cells producing HuMAbs against IL-1 β , TNF- α , IL-6 or IL-10. Detection of IgG autoantibodies to human IL-10 in a human serum is shown in Table 2 below.

Table 2

| Detection of IgG autoantibodies to human IL-10 in human serum | | | |
|---|----------|---|--|
| Sera | Dilution | ¹²⁵ I-IL-10 precipitated with Protein G (cpm) | ¹²⁵ I-IL-10 precipitated after protection with unlabeled IL-10 |
| none | | 405 | 375 |
| negative | 1/2 | 430 | 410 |
| positive | 1/2 | 1425 | 450 |
| | 1/4 | 1142 | 417 |
| | 1/8 | 927 | 391 |
| | 1/16 | 734 | 397 |
| | 1/32 | 442 | 401 |

Generation of X3, a Human Monoclonal Antibody to Human IL-1 α

Plasma from a selected patient (identified as n° X) was found to precipitate human ¹²⁵I-IL-1 α with protein G and inhibited the binding of human ¹²⁵I-IL-1 α to EL4 cells in the protocols described above (see Table 1 above).

EBV Transformation and CD40 Activation of B Lymphocytes from Patient X

40 ml of peripheral blood collected on EDTA treated tubes were obtained from patient X. Blood was diluted 1:1 with PBS and loaded onto a Ficoll® (Pharmacia, Uppsala, Sweden) density gradient. Peripheral blood mononuclear cells (PBMNC) were collected at the interface after centrifugation (30 min, 600 g). Cells were washed four times in PBS, and 39×10^6 PBMNC were finally obtained, with a cell viability superior to 95%, as estimated by Trypan Blue dye exclusion.

The cells were pelleted, resuspended in 1ml of RPMI complete medium which consisted of RPMI 1640 supplemented with 10% heat-inactivated FCS, 2 mM L-glutamine and 50 µg/ml gentamycin. Then, 500 µl of a 100x concentrated Epstein-Barr virus (EBV) suspension (strain B95.8) were added, and this mixture was incubated for 2 hours at 37 °C, 5% CO₂ in humidified incubator. The cells were then washed once in RPMI complete medium and the pellet was resuspended at 5×10^4 cells/ml in Yssel's modified Iscove's medium [Yssel *et al.*, *J. Immunol. Methods* 72 : 219-227 (1984)] supplemented with 15% heat-inactivated FCS, 2 mM L-glutamine, 50 µg/ml gentamycin. Irradiated L cells (7,000 rads) stably expressing the FcγRII (CDw32) were added at a final concentration of 5×10^4 cells/ml, together with the murine monoclonal anti-human CD40 antibody mAb 89 used at a final concentration of 0.5 µg/ml. One hundred µl aliquots of this mixture were then distributed in each well of round-bottomed 96-well culture plates (Nunc), and the plates were incubated at 37 °C, 5% CO₂ atmosphere in humidified incubator. After 5 days of incubation, 125 µl of fresh culture medium containing 0.5 µg/ml anti-CD40 mAb 89 were added to each well.

Screening of Culture Supernatants

After 10 days of incubation, the culture supernatants were screened for the presence of human anti-IL-1α antibodies. Thus, 60 µl of culture medium were collected from each well, and the 12 different supernatants corresponding to the 12 wells of each line of the culture plates were pooled in one microtube. This operation was facilitated by the use of a BIOMEK 1000 work station (Beckman Instruments, Fullerton, CA). Then, 50 µl of the pooled supernatants were screened individually for the presence of antibodies that bind human IL-1α by immunoprecipitation of recombinant human ¹²⁵I-IL-1α with polyspecific anti-human IgM, IgA and IgG antibodies coupled to agarose (Sigma Chemical Co). A total of 13 positive pools were identified with this screening assay. A positive result was immediately confirmed by immunoprecipitation of human ¹²⁵I-IL-1α with protein G coupled to Sepharose, indicating the presence of human IgG antibodies to human IL-1α.

The 13 pools were then identified and split. At day 11, 50 µl of culture supernatants were harvested from each well corresponding to the 13 pools, and they were tested individually by immunoprecipitation of human ¹²⁵I-IL-1α with protein G. A total of 13 different positive wells were thus identified. They were designed X1 to X13.

Characterization of the Human Antibodies to Human IL-1α from the 13 Positive Cell Lines

The 13 initial positive cell lines (X1 to X13) were expanded in order to produce supernatants for further analysis, and the cells were frozen and stored in liquid nitrogen. The positive results of the 13 different lines were verified at different times by immunoprecipitation of human ¹²⁵I-IL-1α with protein G.

Experiments performed with positive culture supernatants have shown that the 13 cell lines secreted human antibodies that inhibit the binding of human ¹²⁵I-IL-1α on EL4 cells.

The isotype of the human anti-IL-1α antibodies contained in these supernatants was determined by using an immunoprecipitation assay of human ¹²⁵I-IL-1α with Affi-Gel 10 beads coated with specific antibodies against human IgG₁, IgG₂, IgG₃ or IgG₄ heavy chain, or antibodies to human kappa (κ) or lambda (λ) light chain. Three of the 13 cell lines secreted a human anti-IL-1α antibody of IgG₁/κ isotype, one was IgG₄/λ and the nine others were IgG₄/κ. In particular, the human anti-IL-1α antibody X3 was of IgG₄/κ isotype.

In a manner similar to the above methods, B cells from 14 other patients that tested positive in the detection screen described above were also transformed with EBV, CD40 activated, screened and expanded. PBLs isolated from these 14 selected patients were submitted to EBV infection, cultured in the CD40 system and screened as described above for patient X. Twenty-eight (28) other B cell lines secreting anti-IL-1α antibodies were identified. Thus, among a total of 482×10^6 PBLs used (from 15 different patients), 41 cell lines secreting anti-human IL-1α antibodies were identified. Forty (40) secreted IgG and 1 secreted IgA antibodies to human IL-1α, which precipitated ¹²⁵I-IL-1α and inhibited its binding on EL4 cells. Details are shown in Table 3 below. While the 40 positive wells other than the one from patient X as described

above lost anti-IL-1 α precipitating activity upon subsequent cloning and subcloning, the B cells in positive wells may be used to identify and isolate other anti-IL-1 α HuMAbs by the other methods described below, e.g., by repertoire cloning.

Table 3

| Generation of anti-IL-1 α secreting human B cell lines | | | | | |
|---|----------------------------------|-----------------------------|-------------------------------|----------------------------|--------------------|
| Patient | N \cdot PBLs ($\times 10^6$) | PBLs/well ($\times 10^3$) | N \cdot positive cell lines | Anti-IL-1 α Isotype | Binding Inhibition |
| C | 50 | 5 | 9 | IgG | + |
| D | 45 | 5 | 1 | IgG | + |
| F | 21 | 5 | 0 | - | - |
| H | 40 | 5 | 1 | IgG | + |
| I | 41 | 5 | 1 | IgG | + |
| P | 90 | 5 | 12 | IgG + IgA | + |
| Q | 20 | 5 | 0 | - | - |
| R | 15 | 4 | 1 | IgG | + |
| S | 40 | 1 | 0 | - | - |
| T | 10 | 1 | 0 | - | - |
| U | 16 | 1 | 0 | - | - |
| V | 15 | 1 | 0 | - | - |
| W | 15 | 1 | 2 | IgG | + |
| X | 39 | 5 | 13 | IgG | + |
| Y | 25 | 5 | 1 | IgG | + |
| TOTAL | 482 | | 41 | 40 IgG + 1 IgA | + |

Cloning of the 13 Positive Cell Lines from Patient X

Twelve days after the initiation of the culture, the 13 positive initial cell lines (X1 to X13) were cloned by limiting dilution at 5 cells/well in 96 round-bottomed microwell plates. Aliquots of cells were harvested, enumerated and resuspended at 50 cells/ml in complete culture medium containing 5×10^4 /ml irradiated (7,000 rads) CDw32 transfected L cells and 0.5 μ g/ml anti-CD40 mAb 89. One hundred μ l of this suspension was distributed in each well and culture plates were incubated at 37 $^\circ$ C, 5% CO $_2$. After 6 days of incubation, 125 μ l of fresh Linolea complete medium were added to each well. Between 10 and 24 days after the cloning initiation, 50 μ l of supernatant were harvested from the wells showing a cell growth, and screened individually for anti-IL-1 α antibodies by the immunoprecipitation assay performed with anti-human IgM, IgA and IgG antibodies-agarose or protein G-Sepharose. The cell line, X3, gave rise to 3 positive clones: X3A, X3B and X3C.

Subcloning of X3A, X3B and X3C

The 3 positive clones X3A, X3B and X3C were then subcloned at 1 cell/well in complete culture medium without feeder cells. The screening was performed at different days after the initiation of the culture, by immunoprecipitation assay as described above. A total of 261 EBV-transformed cell lines secreting human anti-IL-1 α antibodies were obtained. The cells were expanded in RPMI complete medium. Culture supernatants were frozen and stored at -20 $^\circ$ C. Cells were frozen at 1 to 5×10^6 cells/ml and kept in liquid nitrogen.

Antibodies Isotyping in Conditioned Media

Three subclones: X3A-16G5, X3B-14G10 and X3C-20G10, obtained as described above, were selected and maintained in culture in RPMI 1640 supplemented with 10% FCS, 2 mM L-glutamine and 50 μ g/ml gentamycin. Conditioned media from these 3 clones were collected and tested (dilution 1:1 in PBS, 0.05% Tween-20) in enzyme-linked immunosorbent assays (ELISA) specific for human IgM, IgG or IgA isotypes,

and for human IgG₁, IgG₂, IgG₃ or IgG₄ subclasses.

Only a human IgG₄ immunoglobulin has been identified in conditioned media from the three selected subclones X3A-16G5, X3B-14G10 and X3C-20G10, thus suggesting the monoclonality of these three EBV-transformed cell lines. Furthermore, sensitive PCR analysis of V_H, C_H, V_L, and C_L usage indicated the presence of a unique Ig transcript in the isolated B cell clones. The subclone X3A-16G5 was selected for subsequent analysis and is referred to herein as X3.

Purification of the Human Monoclonal Antibody X3

One clone, X3A-16G5, obtained after cloning and subcloning of the EBV-transformed cell line X3, was used to produce large quantities of human monoclonal antibodies to human IL-1 α . This clone was stable for more than 5 months, and was continuously amplified in RPMI 1640 medium supplemented with 10% FCS, 2 mM L-glutamine and 50 μ g/ml gentamycin to allow the generation of large number of cells. Then cells were collected, washed twice in PBS to remove FCS protein contamination, and recultured for 5 days at the initial concentration of 5×10^5 cells/ml in RPMI 1640 supplemented with 2 mM L-glutamine, 50 μ g/ml gentamycin and 1x Nutridoma-HU (Boeringer Mannheim GmbH, Mannheim, Germany). Under these conditions, this clone produced 3 to 5 mg/l of monoclonal IgG₄.

Conditioned culture medium was then collected, filtered and concentrated. Immunoglobulins were then precipitated with 2.4 M (NH₄)₂SO₄. After centrifugation, the precipitate was dissolved in TPP buffer (20 mM H₃PO₄, pH 7) and loaded to an affinity column of protein G-Sepharose 4B (Sigma Chemical Co.) previously equilibrated with TPP buffer. The column was then washed once with TPP buffer, 1 M NaCl, and three times with TPP buffer alone. The human antibody X3 was eluted from the column with 0.1 M glycine buffer, 0.4 M NaCl, pH 2.7. The pH was immediately adjusted to pH 8 by addition of TRIZMA Base 1M, pH 12, and the purified antibody was dialyzed against PBS.

The quality of the X3 purifications was verified by subjecting the obtained preparations to polyacrylamide gel electrophoresis (SDS-PAGE) in a 10% gel under reducing conditions essentially as described by Laemmli [*Nature* 227 : 680-685 (1970)], and after subjecting the gel to silver staining.

To verify that the human antibody X3 was not denatured during the purification, purified preparations of X3 were tested for their ability to precipitate human ¹²⁵I-IL-1 α in the standard assay described above using protein G-Sepharose.

X3 Affinity for Human IL-1 α

To determine the affinity of the human monoclonal antibody X3 for human IL-1 α , the equilibrium association constant of X3/IL-1 α complexes was measured. Purified human antibody X3 was incubated with increasing concentrations of human ¹²⁵I-IL-1 α (10 to 500 pM) in a final volume of 250 μ l in RPMI 1640, 1% BSA, 20 mM Hepes. Each tested condition was performed in triplicate. Nonspecific binding controls were performed in duplicate by addition of 50 nM unlabeled recombinant human IL-1 α . After 4 hours incubation at 4°C, 200 μ l of each sample were distributed in one well of 96 well special titration plates (MultiScreen-HA, 0.45 μ m) containing 50 μ l of protein G coupled to Sepharose. After 1 hour incubation at 4°C, plates were washed four times with PBS and dried membranes were collected from each well. Radioactivity corresponding to the complexes ¹²⁵I-IL-1 α /anti-IL-1 α /protein G-beads retained on the membranes was counted using a Wizard gamma-counter (Wallac). Specific binding of human ¹²⁵I-IL-1 α was calculated, then plotted versus free human ¹²⁵I-IL-1 α concentrations and subjected to Scatchard analysis using a Ligand software (Figure 1).

The equilibrium affinity constant (K_a) value obtained for the human monoclonal antibody X3 was 5×10^9 M⁻¹.

Inhibition of Human IL-1 α Receptor Binding

The ability of human monoclonal antibody X3 to inhibit the binding of radiolabeled human IL-1 α to IL-1 receptors expressed on murine thymoma EL4 cells was investigated using both conditioned medium and the purified antibody. Serial dilutions (in RPMI 1640, 1% BSA, 20mM Hepes) of positive culture supernatants or purified HuMAb X3 were pre-incubated for 1 hour at 4°C with a fixed concentration (70 pM) of human ¹²⁵I-IL-1 α , in a final volume of 100 μ l. Experiments were performed in conical 1 ml Eppendorf tubes or in V-bottomed microtiter plates (Nunc, Roskilde, Denmark), and each sample was tested in triplicate. Then, 1×10^6 (100 μ l) of EL4 cells in RPMI 1640, 1% BSA, 20mM Hepes were added to each tested point and incubated for 3 hours at 4°C, then washed three times, and the radioactivity corresponding to cell-

bound ^{125}I -IL-1 α was counted in a Wizard gamma-counter (Wallac). Non-specific binding was measured in the presence of a 100-fold excess of unlabeled human IL-1 α .

The results presented in Figure 2 indicated that the antibody X3 blocks in a dose-dependent manner the binding of radiolabeled human IL-1 α on EL4 cells. The concentration of antibody X3 required to block 50% of receptor binding (IC_{50}) was found to be 0.015 $\mu\text{g/ml}$ (100 pM) for a constant concentration of 70 pM of radiolabeled human IL-1 α .

Cross-Reactivity of the Human Monoclonal Antibody X3

To determine whether antibody X3 specifically binds to human IL-1 α , different preparations of IL-1 were tested for their ability to protect the precipitation of human ^{125}I -IL-1 α by the human antibody X3. Different dilutions (in PBS, 1% BSA) of purified antibody X3 were preincubated for 1 hour at room temperature without or with an excess (10 nM) of either recombinant human IL-1 α (as positive control), recombinant human IL-1 β or recombinant human IL-1 receptor antagonist (IL-1Ra). Then human ^{125}I -IL-1 α (50 pM) was added to each sample and reaction was incubated for 45 minutes at room temperature. Each sample was tested in duplicate in MultiScreen-HA® plates. Precipitation of ^{125}I -IL-1 α /anti-IL-1 α complexes was done with protein G coupled to Sepharose and radioactivity was counted in a Wallac Wizard gamma-counter.

The results as shown in Figure 3 showed that neither an excess of unlabeled human IL-1 β nor human IL-1Ra protected the immunoprecipitation of human ^{125}I -IL-1 α by antibody X3, while unlabeled human IL-1 α completely inhibited radiolabeled human IL-1 α precipitation. These results indicate that the human monoclonal antibody X3 specifically recognized human IL-1 α , but not human IL-1 β and human IL-1Ra.

Inhibition of Human IL-1 α -Induced IL-2 Secretion by EL4 Cells

The biological activity of human IL-1 was measured by its ability to stimulate IL-2 production by the murine thymoma subline EL4-6.1 [Zubler et al., *J. Immunol.*, 134 : 3662-3668 (1985)]. The IL-2 production was further determined using the CTLL-2 assay [Gillis et al., *J. Immunol.*, 120 : 2027-2033 (1978)]. The proliferation of IL-2 dependent CTLL cells is proportional to the concentration of IL-2 produced by EL4 cells in the first step of culture.

Different concentrations of the purified human monoclonal antibody X3 were incubated for 30 minutes at 37°C with various concentrations of recombinant human IL-1 α or human IL-1 β in a final volume of 100 μl /well in flat-bottomed 96-well culture plates (Falcon, Oxnard, CA). Each experimental point was done in triplicate and reagent dilutions were performed in culture medium composed of RPMI 1640 supplemented with 2 mM L-glutamine, 10% heat-inactivated FCS, 50 $\mu\text{g/ml}$ gentamycin and 5×10^{-5} M 2-mercaptoethanol (2-ME). Then, 100 μl of a suspension of EL4 cells (5×10^5 cells/ml) in culture medium containing 0.2 $\mu\text{g/ml}$ ionomycin (Sigma Chemical Co) were added to each well. After 24 hours incubation at 37°C, 5% CO_2 , in humidified incubator, cell free supernatants were harvested and tested for their IL-2 concentrations.

Aliquots of 50 μl of these supernatants were distributed in flat-bottomed microtiter plates and incubated with 50 μl /well of a suspension of IL-2-dependent CTLL cells (5×10^4 cells/ml) which had been washed twice prior to the assay to remove contamination of IL-2 added to maintain their continuous growth. After 36 hours incubation (37°C, 5% CO_2), cells were pulsed for 4 hours with 0.5 $\mu\text{Ci/well}$ ^3H -Thymidine (specific activity: 25 Ci/mmol, CEA, Saclay, France). ^3H -Thymidine uptake was then measured by standard techniques after harvesting cells on glass fiber filters with a BETAPLATE® 96-well harvester (Pharmacia) and counting in a BETAPLATE® liquid scintillation counter (Wallac). Results were expressed in cpm \pm standard deviation of culture triplicates and are shown in Figure 4.

The human monoclonal antibody X3 specifically inhibits human IL-1 α -induced IL-2 production by EL4 cells, but not the IL-2 secretion induced by human IL-1 β . The concentration of antibody X3 required to block 50% of IL-2 secretion induced by 50 pg/ml (2.8 pM) human IL-1 α (IC_{50}) was found to be 0.1 $\mu\text{g/ml}$ (700 pM).

Inhibition of Human IL-1 α -induced IL-6 Production by Human Synoviocytes

Human synoviocytes were isolated from rheumatoid synovial biopsies obtained from rheumatoid arthritis patients.

Different concentrations of the human monoclonal antibody X3 were incubated for 30 minutes at 37°C with various concentrations of recombinant human IL-1 α or human IL-1 β in a final volume of 100 μl /well in flat-bottomed microtiter plates (Falcon). Each experimental point was done in triplicate and reagent dilutions were performed in culture medium composed of α -MEM (Gibco) supplemented with 2 mM L-glutamine,

10% heat-inactivated FCS, 50 µg/ml gentamycin and 20 mM Hepes. Then, 100 µl/well of a suspension of human synoviocytes (5×10^4 cells/ml in culture medium described above) were added. After 7 days of incubation (37°C, 5% CO₂) supernatants were harvested and tested for their human IL-6 concentrations using a specific ELISA.

Results shown in Figure 5 indicate that the human monoclonal antibody X3 specifically inhibits human IL-1 α -induced IL-6 production by human synoviocytes, but not the IL-6 secretion induced by human IL-1 β . With a constant concentration of 50 pg/ml (2.8 pM) human IL-1 α , the IC₅₀ was found to be 0.02 µg/ml (150 pM) of human monoclonal antibody X3.

Inhibition of Native Human IL-1 α Biological Activity

All the experiments described above were performed using a recombinant form of human IL-1 α . In order to demonstrate that the human antibody X3 also neutralizes the biological activity of native human IL-1 α , we tested the ability of the antibody X3 to inhibit IL-1-related activities contained in conditioned media or lysates of lipopolysaccharide-(LPS) stimulated human mononuclear cells.

To produce native IL-1, PBMNC were isolated from healthy donors by centrifugation on standard Ficoll® density gradient. PBMNC were washed and 10^7 cells were then incubated at 37°C, 5% CO₂ in one well of a 6-well tissue culture plate (Falcon) in 2.5 ml complete culture medium which consisted of RPMI 1640 supplemented with 2 mM L-glutamine, 10% FCS and 50 µg/ml gentamycin. After 1 hour incubation non-adherent cells were removed and the adherent cells were washed three times with culture medium maintained at 37°C. Then adherent cells were re-cultured for 24 hours in 2.5 ml complete culture medium containing 1 µg/ml LPS (*E. coli*, serotype 0111:B4) (Sigma Chemical Co.). After this incubation period, supernatant corresponding to the conditioned medium was harvested and centrifuged to remove contaminating cells. Adherent cells were washed three times with cold PBS and removed by treatment at 4°C with EDTA 0.02% (Sigma) and gentle scrapping with a rubber policeman. Cells were then washed twice, centrifuged, resuspended in 500 µl culture medium and lysed by successive freezings in liquid nitrogen. The cell lysate was then centrifuged (10,000 g, 15 min, 4°C) and the upper phase was collected and adjusted to 1 ml with complete culture medium.

The ability of the human monoclonal antibody X3 to block the native human IL-1 activity contained in conditioned medium or lysate from LPS-activated adherent human mononuclear cells was investigated using the EL4/CTLL assay, because such conditioned medium and lysate may contain significant amounts of LPS and IL-6 which may interact in the synoviocyte assay, but not in the EL4/CTLL assay.

Serial dilutions of conditioned medium or lysate from LPS-activated adherent cells were preincubated for 30 minutes without or with 1 µg/ml purified antibody X3, 1 µg/ml non-related IgG₄/x human monoclonal antibody (as negative control) or 1 µg/ml rabbit neutralizing antibodies against human IL-1 α (Genzyme) (as positive control). Each condition was done in triplicate, in 96-flat-bottomed culture plates under a final volume of 100 µl/well. Then 100 µl/well of EL4 cells (5×10^5 cells/ml) were added and incubated 24 hours at 37°C, 5% CO₂ in humidified incubator. Culture supernatants were then collected and their IL-2 concentrations were determined using the CTLL assay.

Results presented in Table 4 below show that the human antibody X3 inhibited the IL-1 activity contained in lysate but not in conditioned medium obtained from LPS-activated human mononuclear cells. The neutralizing rabbit anti-human IL-1 α antiserum shared the same activities as antibody X3, while the unrelated human antibody did not inhibit the IL-1 activity neither in conditioned medium, nor in cell lysate. These results indicate that the antibody X3 recognizes and neutralizes human native IL-1 α (which is mostly present into the cytosol and associated to the membrane of LPS-activated monocytes), but not native IL-1 β (which is principally secreted after LPS activation).

Table 4

| Inhibition of native human IL-1 by antibody X3 | | | | | |
|--|----------|--|-----------------------------|------|---------------------------|
| LPS-activated adherent MNC | | EL4/CTLL proliferation [³ H]Thymidine uptake (cpm x 10 ⁻³) | | | |
| Preparation | Dilution | Medium | Control IgG ₄ /x | X3 | Rabbit anti-IL-1 α |
| Lysate | 1/40 | 84.4 | 115.4 | 18.2 | 22.2 |
| | 1/400 | 26.8 | 35.7 | 2.9 | 4.2 |
| Supernatant | 1/400 | 85.8 | 98.0 | 82.9 | 89.4 |
| | 1/4000 | 5.5 | 4.9 | 5.1 | 4.2 |

The reactivity of HuMAb X3 with Cynomolgus IL-1 was also tested. Monkey blood mononuclear cells were isolated, stimulated with LPS and lysed after 24 hours incubation. Increasing concentrations of lysate induced EL-4 cells to secrete IL-2 and this activity was inhibited by the polyclonal rabbit anti-human IL-1 α . HuMAb X3 was also able to block the monkey IL-1 α , but its activity appeared to be lower than that observed with human IL-1 α .

20 Inhibition of Membrane Associated Human IL-1 α Activity

The reactivity of X3 with membrane associated IL-1 α was studied by using highly purified human monocytes metabolically inactivated after paraformaldehyde (PFA) fixation. Monocytes were isolated from peripheral blood by elutriation centrifugation essentially as described by De Mulder et al., *J. Immunol. Methods*, 47 : 31-38 (1981). Purity of the different fractions obtained was assessed by flow cytometry analysis, and preparations containing more than 90% CD14 positive cells were selected. Monocytes were then cultured with or without LPS (1 μ g/ml) in teflon cell culture bags for 24 hours at 37°C, 5% CO₂. Then cells were washed twice in PBS and resuspended in PBS containing 1% PFA (Sigma) for 10 minutes at 20°C. Cells were then washed three times with glycine buffer (150 mM glycine, 75 mM NaCl, pH 7.4) and three times with RPMI 1640 complete culture medium. Serial dilutions of PFA-fixed monocytes were then incubated 30 minutes at 37°C in a final volume of 100 μ l/well with or without either 1 or 10 μ g/ml monoclonal antibody X3, or 10 μ g/ml rabbit anti-IL-1 α or anti-IL-1 β antiserum. Cultures were performed in flat-bottomed 96-well culture plates. Then 100 μ l of EL4 suspension (5x10⁵ cells/ml) were added to each well. After 48 hour incubation IL-2 secretion was measured with the CTLL-2 assay as previously described. Results are presented in Figure 6.

The human monoclonal antibody X3 was found to inhibit IL-2 secretion by EL4 cells induced by PFA-fixed monocytes activated or not with LPS. A similar inhibition was obtained with the rabbit anti-IL-1 α antibody but not with the rabbit anti-IL-1 β antiserum. These results indicate that the HuMAb X3 recognizes and neutralizes human membrane IL-1 α .

40 Inhibition of IL-6 Production In Cocultures of Synoviocytes and Monocytes

To test the effect of HuMAb X3 on the production of IL-6 by the coculture of synoviocytes and monocytes, elutriated blood monocytes were cultured for 24 hours with or without LPS (1 μ g/ml) and then fixed or not with PFA as described above. Serial dilutions of monocyte preparations were then incubated for 30 minutes at 37°C with or without 1 μ g/ml of HuMAb X3, 1 μ g/ml non-related human IgG₄ antibody or 100 ng/ml of IL-1Ra. Cultures were performed in triplicates, in flat-bottomed 96-well culture plates. Then 100 μ l of synoviocyte suspension (5x10⁴ cells/ml) were added to each well. After 48 hours of incubation, IL-6 secretion was measured in supernatants with a specific ELISA. Controls performed without synoviocytes showed that, in contrast to unfixed monocytes, PFA-fixed monocytes were unable to secrete IL-6. The results are shown in Figure 7.

The rheumatoid synovial tissue is composed of about 20% monocyte/macrophage/dendritic cells, 20% fibroblast-like cells (synoviocytes) and 30-50% T cells. This inflammatory tissue produces *in vivo* and *ex vivo* high levels of proinflammatory cytokines, including IL-6, TNF- α and IL-1 β , Miossec et al., *Arthritis Rheum.*, 35 : 874-883 (1992). A coculture of freshly isolated monocytes with synoviocytes from long term cultures results in the production of large amounts of IL-6 but not of IL-10, IL-1 β or TNF- α . Furthermore, a coculture of synoviocytes with PFA-fixed monocytes (unable to secrete IL-6) also produced large amounts of IL-6. This indicates that IL-6 is most likely produced by synoviocytes following contact with monocytes.

As shown in Figure 7, HuMAb X3 was found to strongly inhibit the production of IL-6 by coculture of synoviocytes with non-activated monocytes or LPS-activated monocytes (without or with PFA fixation). This finding is in accordance with the inhibitory effect of IL-1Ra. Thus, HuMAb X3 is able to interrupt an interaction between monocytes and synoviocytes which may represent a critical step in the development of rheumatoid inflammation.

Sequencing of the Variable Region Genes of the Anti-IL-1 α HuMAb X3

RNA from the B cell clone X3A-16G5 has been isolated according to the single-step method described by Chomczynski, et al., *Anal Biochem*, 162 : 156-9 (1987). Briefly, about 10^7 cells from this clone were lysed in guanidinium thiocyanate denaturing solution. After acidification of the mixture with 2M sodium acetate, pH4, RNA was extracted with phenol and chloroform/isoamyl alcohol (24:1). RNA was then precipitated with isopropanol, the RNA pellet was redissolved in denaturing solution, reprecipitated with isopropanol, and washed with 75% ethanol.

cDNA was obtained by reverse transcription, using the Superscript Reverse Transcriptase Kit (cat. 20898 BRL, Gaithersburg, MD, USA), with oligo dT₁₂₋₁₈ primers (Cat. 27.7858-01, Pharmacia, Uppsala, Sweden). The cDNA was then used as template in the polymerase chain reaction (PCR). PCR amplifications were performed with Taq polymerase (Perkin Elmer, Norwalk, Connecticut) using the reaction buffer provided by the manufacturers: Taq buffer: 1,5 mM MgCl₂, 50 mM KCl, 10 mM Tris-HCl, pH 8.3 and 0.001% (w/v) gelatin. All PCR reaction mixture contained 200 ng of each primer, and 2.5 of Taq Polymerase. Amplifications were performed in a Trio-Thermoblock Thermal cycler (Biometra, GmbH) and consisted of 35 cycles of 1 minute denaturation at 94 °C, 2 min of primer annealing at 60 °C, and 3 minutes extension at 72 °C. After the last cycle, the reaction mixtures were incubated for 10 minutes at 72 °C to insure complete extension of all products. The primers were designed to include restriction sites, to allow for the directional cloning of the PCR products. For the heavy chain, primers (listed in SEQ ID NOS. 3, 4, 5, 6, 7 and 8) specific for the leader sequence of the 6 different human VH families were used individually in conjunction with a primer (listed in SEQ ID NO. 9) located at the 3' end of the gamma constant region corresponding to the sub-class previously determined by isotyping the HuMAb by ELISA (IgG4). The light chain was amplified with individual combinations of primers corresponding to the 3' end of the kappa or lambda chain (listed in SEQ ID NOS. 10 and 11, respectively) in conjunction with a series of primers (listed in SEQ ID NOS. 12, 13, 14, 15, 16 and 17) annealing to the leader sequence of the different V kappa gene families or with a series of primers (listed in SEQ ID NOS. 18, 19, 20, 21 and 22) annealing to the leader sequence of the different V lambda gene families. Thus, full length heavy and light chains starting at the initiation codon in the leader sequence and ending at the stop codon have been generated. Two independent PCR were performed for both the heavy and the light chains.

Those PCR products were loaded on agarose gels, and purified with GELase (Epicentre, cat. G21223, WI) according to the manufacturer's instructions. Purified PCR products from heavy and light chains were used as template for sequencing reaction with leader PCR primers and with primer hybridizing at the 5' end of the gamma and of the kappa or lambda constant region gene respectively. The sequencing reaction was performed on a 373 DNA Sequencer with TaqDyeDeoxy Terminator Cycle Sequencing Kit (both from Applied Biosystems Inc. Foster City, CA). Direct sequencing of both strands of the products of 2 independent PCRs were therefore obtained and compared. No difference was found between the sequences of the 2 PCRs from the same cells. The sequence of the V_H gene is listed in SEQ ID NO. 1 and the sequence of the V_L gene is listed in SEQ ID NO. 2 below. Framework (FR) and complementarity-determining segments (CDR) are identified as below, in agreement with the system of Kabat et al. (Kabat, E.A., T.T. Wu, H.M. Perry, K.S. Gottesman, and C. Foeller. 1987. *Sequences of Proteins of Immunological Interest*. United States Department of Health and Human Services, Bethesda, MD, p. 1.).

| V _H Segment - SEQ ID NO. 1 | |
|---------------------------------------|----------------|
| amino acid residue nos. | region |
| -1 to -18 | Signal peptide |
| + 1 to +30 | FR 1 |
| +31 to +35 | CDR 1 |
| +36 to +49 | FR 2 |
| +50 to +66 | CDR 2 |
| +67 to 98 | FR 3 |
| +99 to +110 | CDR 3 |
| +111 to +122 | JH 1 |

| V _L Segment - SEQ ID NO. 2 | |
|---------------------------------------|----------------|
| amino acid residue nos. | region |
| -1 to -22 | Signal peptide |
| + 1 to +23 | FR 1 |
| +24 to +34 | CDR 1 |
| +35 to +49 | FR 2 |
| +50 to +56 | CDR 2 |
| +57 to 88 | FR 3 |
| +89 to +95 | CDR 3 |
| +96 to +108 | JK 4 |

Sequences were compared with the Gene Bank release 77 using the DNASTar (WI. USA) software:

For the heavy chain sequence (SEQ ID NO. 1), the most homologous sequence found was HUMIGHYAAG, a human VH3 germline gene [Olee, et al., *J. Clin. Invest.*, 88 : 193-203 (1991)]. However, the level of homology was only 91.6% (25 nucleotide mismatches out of 300 nucleotides) suggesting that the germline counterpart of HuMAb X3 heavy chain is probably different from HUMIGHYAAG. It is therefore currently impossible to assign mutation to any of the mismatches observed in the V_H segment. D segments are not unambiguously identifiable. There is one replacement mutation in the JH1 segment (120 Val/Phe).

For the light chain sequence (SEQ ID NO. 2), the most homologous sequence found was HUMIGKVJ2, a human Vkl germline gene [Pech, et al., *J. Mol. Biol.*, 176 : 189-204 (1984)]. The level of homology reaches 94.7% (14 nucleotide mismatches out of 264 nucleotides) strongly suggesting that the germline counterpart of HuMAb X3 light chain is indeed HUMIGKVJ2. Analysis of somatic mutations in the V_L segment showed the following: the ratio of replacement mutations (R) vs. silent mutations (S) is R/S = 5/1 in the CDRs, while R/S is 1/7 in the framework segments (FRs). D segments are not unambiguously identifiable. There are 3 replacement and one silent mutations in the JK4. In conclusion, the kappa light chain of HuMAb X3 is heavily mutated, and the ratio of R/S mutations in the CDRs vs FRs suggests strong selection by the antigen.

45 **Expression of Recombinant Anti-IL-1 α HuMAb In Baculovirus**

RNA from the B cell clone X3A-16G5 was isolated by the guanidinium thiocyanate single-step method described by Chomczynski et al., *supra*. cDNA was obtained by reverse transcription of the RNA using a Superscript Reverse Transcriptase Kit with oligo dT₁₂₋₁₈ primers (Cat. 27.7858-01, Pharmacia, Upsalla, Sweden). The cDNA was then used as a template in the polymerase chain reaction (PCR) performed with Taq polymerase. The primers were designed to include *Eco*R1 and *Not*I restriction sites, to allow for the directional cloning of the PCR products into the baculovirus vector pVL-1393. For the heavy chain, primer (listed in SEQ ID NO. 5) specific for the leader 3 sequence of the human V_H family was used in conjunction with a primer (listed in SEQ ID NO. 9) located at the 3' end of the gamma constant region corresponding to the sub-class previously determined by isotyping the HuMAb by ELISA (IgG₄). The light chain was amplified with a primer (listed in SEQ ID NO. 10) corresponding to the 3' end of the kappa chain in conjunction with a primer (listed in SEQ ID NO. 12) annealing to the leader sequence of the V kappa 1 gene family. Thus, full length heavy and light chains cDNAs were generated. Two independent PCRs were

performed for both the heavy and the light chains.

After appropriate restriction cleavage of these PCR products, both full length heavy and light chains were cloned in baculovirus vector restricted with the same enzymes. Heavy and light chains were cloned individually in distinct pVL1393 baculovirus vectors (Invitrogen Co, San Diego, CA). The recombinant
 5 vectors were transfected in competent DH5 α *E coli* bacteria (Gibco BRL, Gaithersburg, MD), and 10 single colonies were selected. One hundred ml culture of each bacterial clone were obtained and vector DNA was purified with Qiagen plasmid-Kit (Qiagen, GmbH). Both strands of the complete insert from double-stranded DNA vector were sequenced with (1) two primers flanking the insert - the first (listed in SEQ ID NO. 23) annealing 5' in the promoter region of the polyhedrin gene and the second (listed in SEQ ID NO. 24)
 10 annealing 3' in the polyhedrin gene itself; and (2) a series of forward and backward primers distributed about 400 bp apart along the heavy and the light chain sequences, i.e., the forward primers for the heavy chain are listed in SEQ ID NOS. 25 and 26, the backward primers for the heavy chain are listed in SEQ ID NOS. 27, 28 and 29, and the backward primer for the light chain is listed in SEQ ID NO. 30. Double
 15 stranded DNA sequencing was done on a 373 DNA Sequencer with TaqDyeDeoxy Terminator Cycle Sequencing Kit (both from Applied Biosystems Inc., Foster City, CA). A recombinant baculovirus vector clone was selected for both heavy and light chains, which showed perfect match with the variable region sequences obtained from the PCR products, and with the published sequences of the constant regions of the heavy gamma 4 and kappa light chains respectively. Recombinant baculovirus vectors were cotransfected with wild type baculovirus DNA in Sf9 insect cells, using the transfection module (Invitrogen Co, San
 20 Diego, CA). Recombinant baculoviruses recovered from the cell culture supernatant of these transfected cells were then cloned in Sf9 cells by limiting dilution and screened by hybridization with the labeled inserts. After two runs of cloning, followed by production, recombinant baculoviruses containing the heavy or the light chains cDNAs were titrated, and used to infect insect cells at a Multiplicity of Infection (MOI) of 5. After 5 days of culture, production of human heavy or light chain was confirmed by ELISA and/or by in
 25 vivo labeling. One baculovirus clone expressing the heavy chain and one expressing the light chain were used to co-infect Sf9 cells both at a MOI of 5. After 5 days of infection, the presence in the supernatant of an antibody binding specifically to human IL-1 α was confirmed by immunoprecipitation.

Supernatants of insect cells infected with (1) recombinant baculovirus containing cDNA of the light chain
 30 of X3, (2) with baculovirus containing cDNA of X3 heavy chain, (3) with baculoviruses containing cDNAs of both the X3 heavy and the X3 light chains and (4) with baculoviruses containing cDNAs of both heavy and light chains of a non-related human IgG antibody, were assayed in the Standard Immunoprecipitation Protocol with Protein G described above. The results are shown in Figure 8A. The results demonstrate that supernatants containing the X3 heavy and light chains precipitated ¹²⁵I-IL-1 α , while supernatants containing
 35 X3 light chain alone, X3 heavy chain alone, or the heavy and light chains of the non-related human IgG did not immunoprecipitate ¹²⁵I-IL-1 α .

Furthermore, the natural HuMAb X3 and the recombinant form of X3 were employed in the Cross-Reactivity of the Human Monoclonal Antibody X3 assay described above. The results are shown in Figure 8B, which demonstrate that, as with the natural HuMAb X3, the recombinant form of X3 specifically
 40 recognized human IL-1 α , as the ¹²⁵I-IL-1 α immunoprecipitation was not protected by preincubation of X3 with an excess of unlabeled human IL-1 β and IL-1Ra.

Finally, the supernatant of insect cells expressing recombinant HuMAb X3 inhibited IL-1 α biological activity measured in the EL4/CTLL assay described previously, but not the activity of IL-1 β .

45 Repertoire Cloning

As demonstrated above, by dilution cloning of an immortalized and/or activated B cell population in accordance with the present invention, a series of amplified B cell subpopulations can be provided for screening for antibodies that bind to the desired antigen, e.g., by the standard and derived im-
 50 munoprecipitation protocols described above. Thus, by using the immortalization, amplification and screening techniques of the present invention, it is possible to produce and identify an immortalized and/or secreting B cell subpopulation that consists of from about 5 to about 50 different, amplified B cell clones, at least one of which expresses a HuMAb against the desired antigen, e.g. a human cytokine such as IL-1 β , TNF- α , IL-6, IL-10 etc.

For example, as discussed above, we produced amplified, immortalized and/or secreting B cell subpopulations from 15 patients expressing HuMAbs against human IL-1 α . Each of these subpopulations contained from about 5 to about 50 different amplified B cell clones in a total number of about 5x10⁵ to about 50x10⁵ B cells.

Thus, the percentage of B cells producing HuMAbs against the antigen of interest in a B cell subpopulation of the present invention is greatly enhanced in comparison to other techniques which start with naturally occurring B cell populations. This amplified subpopulation can make it possible to uncover HuMAbs to human cytokines even when it is not possible to isolate a single clone as we accomplished with the X3A-16G5 clone discussed above.

Starting with an amplified, immortalized and/or secreting B cell subpopulation in accordance with the present invention, including, for example, 40 different B cell clones, the number of possible V_H/V_L combinations from a cDNA library encoding the V_H segments and V_L segments from these B cells is 1600 (40 x 40). This very low number makes it far easier to isolate the specific combination of V_H and V_L segments responsible for the one (or more) amplified HuMAb clone in the subpopulation which binds to the desired antigen. Thus, by applying standard techniques such as repertoire cloning and phage display to the amplified, immortalized and/or secreting B cell subpopulation of the invention, a series of HuMAbs against the desired antigen can be identified and isolated by recombinant techniques.

For example, a cDNA library encoding the mRNA repertoire of V_H and/or V_L segments of all the HuMAbs expressed in such a subpopulation (e.g., a subpopulation screened as containing a clone expressing a HuMAb to IL-10) can be prepared by PCR amplification of the mRNA from the subpopulation using appropriate primers. These repertoire cloning techniques are now standard in the art, see, for example, Marks et al., *J. Mol. Biol.*, 222 : 581-597 (1991); Huse, et al., *Science*, 246 : 1275-1281 (1989); WO 90/14430; WO 92/15678; WO 91/16427; and WO 92/01047. The DNAs encoding the V_H and V_L segments may be assembled into appropriate vectors for direct cloning and expression in a host, e.g., by the methods described in Hoogenboom, et al., *Nucleic Acids Research*, 19 : 4133-4137 (1991). The expressed V_H and V_L segments or Fab (e.g., for an anti-IL-10 HuMAb) may then be screened for binding to the desired antigen by the standard and derived immunoprecipitation protocols described above using labeled antigen, e.g. 125 I-IL-10. The DNA encoding the V_H or V_L segments from the identified clones can then be sequenced and operatively linked to DNA segments encoding the constant regions for the desired HuMAb isotype heavy or light chains, e.g., IgG₁, IgG₂, IgG₃, IgG₄, IgA, etc. heavy chains or κ or λ light chains, to create a complete HuMAb or a fragment thereof (e.g., Fab, F(ab')₂, Fv etc.) against the desired antigen, e.g., human IL-10.

Alternatively, the cDNA repertoire encoding the V_H and/or V_L segments can be included in a vector appropriate to display the V_H and/or V_L segments on the surface of a suitable host. See, for example, Marks, et al., *J. Mol. Biol.*, 222 : 581-597 (1991) and Hoogenboom et al., *supra*, which disclose methodologies for displaying Fv, single chain Fv (scFv) or Fab fragments of such a cDNA library on the surface of bacteriophage. The host cells (e.g., phage) displaying the scFv on their surface that bind to the desired antigen can then be identified by ELISA or any other suitable assay. The DNA encoding the V_H and/or V_L segments from binding host cells can be separated and reassembled into appropriate vectors for direct cloning and expression in a host, e.g., by the method described in the Marks et al. article. The DNA sequences can then be assembled into a full length HuMAb or fragment thereof by the methodologies described above.

An amplified B-cell subpopulation of the present invention as described above may be employed with single cell and multiple cell PCR techniques to obtain the DNA sequences encoding the variable regions of the HuMAbs produced by these cells. For example, a CD40 crosslinked B-cell population of the invention (which may also be EBV-transformed) may be diluted to provide either a small number of B-cells (e.g., 10 B-cells per well) or a single B-cell or less per well. The methodologies disclosed, for example, in Larrick et al., *Biotechnology*, 7 : 934 (1989), Embleton et al., *Nucleic Acids Research*, 10 : 3831-3837 (1992), Liu et al., *Proc. Natl. Acad. Sci.*, 89 : 7610-7614 (1992) and Lew et al. *Immunology*, 75 : 3-9 (1992) may be employed to obtain accurate and complete heavy and light chain variable region (V_H and V_L) genes from these cells. These DNA sequences may be included in an appropriate recombinant system to express, for example, an Fv or ScFv, and the fact that they represent a HuMAb to the desired antigen may then be confirmed by the immunoprecipitation assays described above for binding to the desired antigen. Previous isotyping of the active HuMAbs in the B-cell starting population or subpopulation may then be used to construct the full length HuMAb.

The activity of the HuMAb may then be confirmed by conventional *in vitro* and *in vivo* biological assays. For example, for TNF- α , IL-1 β and IL-6, the assays and models reviewed in Dinarello, *Eur. Cytokine Netw.*, 3 : 7-17 (1992) may be employed. For human IL-10 the cytokine synthesis inhibitory factor (CSIF) assay described in Fiorentino et al., *J. Exp. Med.*, 170 : 2081-2095 (1989) or the property of IL-10 to induce proliferation and Ig secretion by human B cells as described in Rousset et al., *Proc. Natl. Acad. Sci. USA*, 89 : 1890-1893 (1992) may be employed.

It is most likely that the pairs of heavy and light chains identified under the above conditions will be those of the identified antibody as the screening procedures described above are selective for high affinity antibody which can only be obtained with a given combination of heavy and light chain. Furthermore, the identification of the heavy and light chain isotypes in the supernatants of the oligoclonal cell lines will also be of great help to determine whether the selected pair does indeed correspond to the initially identified clone.

While the present invention has been described in conjunction with the specific embodiments set forth above, many alternatives, modifications and variations thereof will be apparent to those of ordinary skill in the art. All such alternatives, modifications and variations fall within the spirit and scope of the present invention.

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SEQUENCE LISTING

5 (I) GENERAL INFORMATION:

(i) APPLICANT:

10 (A) NAME: SCHERING-PLOUGH

(B) STREET: 92 RUE BAUDIN

15 (C) CITY: 92307 LEVALLOIS-PERRET, Cedex

20 (D) STATE:

(E) COUNTRY: France

25 (F) POSTAL CODE: 92307

30 (ii) TITLE OF INVENTION: Human Monoclonal Antibodies
Against Human Cytokines And Methods Of Making And
Using Such Antibodies

35 (iii) NUMBER OF SEQUENCES: 30

(v) COMPUTER READABLE FORM:

40 (A) MEDIUM TYPE: Floppy disk

(B) COMPUTER: Apple Macintosh

45 (C) OPERATING SYSTEM: Macintosh 6.0.8

(D) SOFTWARE: Microsoft Word 5.1a

50

55

(2) INFORMATION FOR SEQ ID NO: 1:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 423 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

ATG GAG TTT GGG CTG AGC TGG GTT TTC CTC GTT GCT CTT TTA AGA GGT 48
 Met Glu Phe Gly Leu Ser Trp Val Phe Leu Val Ala Leu Leu Arg Gly

-15

-10

-5

GTC CAG TGT CAG GTG CAA CTG GTG GAA TCT GGG GGA GGC GTG GTC CAG 96
 Val Gln Cys Gln Val Gln Leu Val Glu Ser Gly Gly Gly Val Val Gln

+1

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CCT GGG AGG TCC CTG AGA CTC TCC TGT ACA GCC TCT GGA TTC ACC TTC 144
 Pro Gly Arg Ser Leu Arg Leu Ser Cys Thr Ala Ser Gly Phe Thr Phe

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AGT ATG TTT GGT GTC CAC TGG GTC CGC CAG GCC CCA GGC AAG GGG CTG 192
 Ser Met Phe Gly Val His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu

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GAG TGG GTG GCA GCT GTG TCA TAT GAT GGA AGC AAT AAG TAC TAT GCA 240
 Glu Trp Val Ala Ala Val Ser Tyr Asp Gly Ser Asn Lys Tyr Tyr Ala

50

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60

5 65 70 75
 GAG TCC GTG AAG GGC CGA TTC ACC ATC TCC AGA GAC AAT TCC AAG AAC 288
 Glu Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn
 10 80 85 90
 ATC TTA TTT CTA CAA ATG GAC AGC CTG AGA CTT GAG GAC ACG GCT GTC 336
 Ile Leu Phe Leu Gln Met Asp Ser Leu Arg Leu Glu Asp Thr Ala Val
 15 95 100 105
 TAT TAC TGT GCT AGA GGC CGG CCC AAG GTC GTA ATA CCA GCA CCT TTG 384
 Tyr Tyr Cys Ala Arg Gly Arg Pro Lys Val Val Ile Pro Ala Pro Leu
 20 110 115 120
 GCT CAC TGG GGC CAG GGA ACC CTG GTC ACC TTC TCC TCA 423
 Ala His Trp Gly Gln Gly Thr Leu Val Thr Phe Ser Ser

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 390 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

45 -20 -15 -10
 ATG GAC ATG AGG GTC CCC GCT CAG CTC CTG GGG CTC CTG CTG CTC TGG 48
 Met Asp Met Arg Val Pro Ala Gln Leu Leu Gly Leu Leu Leu Leu Trp
 50

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EP 0 659 766 A1

5 TTC CCA GGT TCC AGA TGC GAC ATC CAG ATG ACC CAG TCT CCA TCT TCC 96
 Phe Pro Gly Ser Arg Cys Asp Ile Gln Met Thr Gln Ser Pro Ser Ser
 -5 +1 5 10

10 GTG TCT GCA TCT GTA GGA GAC AGA GTC ACC ATC ACT TGT CGG GCG AGT 144
 Val Ser Ala Ser Val Gly Asp Arg Val Thr Ile Thr Cys Arg Ala Ser
 15 20 25

15 CAG GGT ATT AGC AGT TGG TTA GCC TGG TAT CAG CAG AAA CCA GGA AAG 192
 Gln Gly Ile Ser Ser Trp Leu Ala Trp Tyr Gln Gln Lys Pro Gly Lys
 30 35 40

20 GCC CCG AAG CTC TTG ATC TAT GAA GCA TCC AAT TTG GAA ACT GGG GTC 240
 Ala Pro Lys Leu Leu Ile Tyr Glu Ala Ser Asn Leu Glu Thr Gly Val
 45 50 55

25 CCA TCA AGA TTC AGC GGC AGT GGA TCT GGG TCA GAT TTC ACC CTC ACC 288
 Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Ser Asp Phe Thr Leu Thr
 60 65 70

30 ATC AGC AGC CTG CAG CCT GAA GAT TTT GCA ACT TAT TAT TGT CAA CAG 336
 Ile Ser Ser Leu Gln Pro Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln
 75 80 85 90

35 ACT AGC AGT TTT CTC CTC AGT TTC GGC GGC GGG ACC AAG GTG GAG CAC 384
 Thr Ser Ser Phe Leu Leu Ser Phe Gly Gly Gly Thr Lys Val Glu His
 95 100 105

40 AAA CGA 390
 Lys Arg

45

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

CCCGAATTCA TGGACTGGAC CTGGAGG 27

(2) INFORMATION FOR SEQ ID NO: 4:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 29 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

CCCGAATTCA TGGACATACT TTGTACCAC 29

(2) INFORMATION FOR SEQ ID NO: 5:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

CCCGAATTCA TGGAGTTTGG GCTGAGC 27

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 29 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

CCCGAATTCA TGAAACACCT GTGGTTCTT 29

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 29 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

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CCCGAATTCA TGGGGTCAAC CGCCATCCT 29

10

(2) INFORMATION FOR SEQ ID NO: 8:

(1) SEQUENCE CHARACTERISTICS:

15

(A) LENGTH: 29 base pairs

20

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

30

CCCGAATTCA TGTCTGTCTC CTCCTCAT 29

35

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

40

(A) LENGTH: 31 base pairs

45

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

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GAGAGAGCGG CCGCACTCAT TTACCCGGAG A 31

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(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 34 base pairs

15

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

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GAGAGAGCGG CCGCTAACAC TCTCCCCTGT TGAA 34

30

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 38 base pairs

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(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

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GAGAGAGCGG CCGCCTATGA ACATTCTGTA GGGGCCAC 38

55

(2) INFORMATION FOR SEQ ID NO: 12:

5 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 36 base pairs

10 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

15 (D) TOPOLOGY: linear

20 CCCGAATTCA TGGACATGAG GGTCCCCGCT CAGCTC 36

25 (2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 33 base pairs

(B) TYPE: nucleic acid

35 (C) STRANDEDNESS: single

40 (D) TOPOLOGY: linear

45 CCCGAATTCA TGGACACGAG GGCCCCCACT CAG 33

(2) INFORMATION FOR SEQ ID NO: 14:

50 (i) SEQUENCE CHARACTERISTICS:

55

(A) LENGTH: 29 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

CCCGAATTCA TGGTGTTCGA GACCCAGGT 29

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 33 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

CCCGAATTCA TGAGGCTCCC TGCTCAGCTC CTG 33

(2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 29 base pairs

(B) TYPE: nucleic acid

5 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

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CCCGAATTCA TGGAAACCCC AGCGCAGCT 29

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(2) INFORMATION FOR SEQ ID NO: 17:

20 (1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 30 base pairs

25 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

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CCCGAATTCA TGGGGTCCCA GGTTCACCTC 30

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(2) INFORMATION FOR SEQ ID NO: 18:

(1) SEQUENCE CHARACTERISTICS:

45 (A) LENGTH: 29 base pairs

(B) TYPE: nucleic acid

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(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

5

CCCGAATTCA TGACCTGCTC CCCTCTCCT 29

10

(2) INFORMATION FOR SEQ ID NO: 19:

(1) SEQUENCE CHARACTERISTICS:

15

(A) LENGTH: 33 base pairs

20

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

25

(D) TOPOLOGY: linear

30

CCCGAATTCA TGGCCTGGAC TCCTCTCTTT CTG 33

35

(2) INFORMATION FOR SEQ ID NO: 20:

(1) SEQUENCE CHARACTERISTICS:

40

(A) LENGTH: 29 base pairs

45

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

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CCCGAATTCA TGGCCTGGGC TCCACTACT 29

5

(2) INFORMATION FOR SEQ ID NO: 21:

10

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 29 base pairs

15

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

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CCCGAATTCA TGGCATGGAT CCCTCTCTT 29

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(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

35

(A) LENGTH: 30 base pairs

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(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

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CCCGAATTCA TGGCCTGGGC TCTGCTGCTC 30

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(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

ACCTATAAAT ATTCCGGATT ATTCA 25

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

TCTTGCCGGG TCCCAGG 17

(2) INFORMATION FOR SEQ ID NO: 25:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 21 base pairs
- (B) TYPE: nucleic acid
- 10 (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- 15

GGTCTCCAAC AAAGCCCTCC C 21

20

(2) INFORMATION FOR SEQ ID NO: 26:

25 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
- 30 (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- 35 (D) TOPOLOGY: linear

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CACAAGCCCA GCAACACCAA GGTGGAC 27

45

(2) INFORMATION FOR SEQ ID NO: 27:

(i) SEQUENCE CHARACTERISTICS:

- 50 (A) LENGTH: 26 base pairs

55

(B) TYPE: nucleic acid

5

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

10

GGGGGGAAGA GGAAGACTGA CGGTCC 26

15

(2) INFORMATION FOR SEQ ID NO: 28:

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 28 base pairs

25

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

30

(D) TOPOLOGY: linear

35

GGGTGTACAC CTGTGGTTCT CCGGGCTG 28

40

(2) INFORMATION FOR SEQ ID NO: 29:

(i) SEQUENCE CHARACTERISTICS:

45

(A) LENGTH: 21 base pairs

50

(B) TYPE: nucleic acid

55

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

GCAGGTGTAG GTCTGGGTGC C 21

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

TGGCGGAAG ATGAAGACAG 20

Claims

1. A human monoclonal antibody against a human cytokine or a fragment of such an antibody, said fragment having an affinity for said human cytokine of 10^8 M⁻¹ or greater.
2. The antibody or fragment according to claim 1 which binds to a human lymphokine or a human monokine.
3. The antibody or fragment according to claim 1 which binds to a human interleukin.
4. The antibody or fragment according to claim 1 which binds to human IL-1 α .
5. The antibody according to any one of claims 1-4 which has an affinity of greater than 10^9 M⁻¹.
6. The antibody according to any one of claims 1-5 which is of the IgG class.
7. A human monoclonal antibody or a fragment thereof characterized in that it comprises at least one CDR of an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 and/or of an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2.; or one or more somatic variant of such sequence.

8. A human monoclonal antibody or a human IL-1 α binding fragment thereof characterized in that it comprises
 - a V_H segment having an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 or by a CDR somatic variant thereof, and/or
 - 5 a V_L segment having an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2 or by a CDR somatic variant thereof.
9. The antibody according to claim 8, characterized in that it comprises a V_H segment having an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 and/or a V_L segment having an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2.
10. The antibody according to claim 8, characterized in that it comprises V_H and V_L segments having the amino acid sequences defined by amino acid residues 1-122 of SEQ ID NO. 1 and amino acid residues 1-108 of SEQ ID NO. 2, respectively, or which comprises a CDR somatic variant of one or both of said amino acid sequences.
11. The antibody according to claim 8, characterized in that it comprises V_H and V_L segments having the amino acid sequences defined by amino acid residues 1-122 of SEQ ID NO. 1 and amino acid residues 1-108 of SEQ ID NO. 2, respectively.
12. The antibody according to claim 10 or 11, characterized in that it is of the IgG₄ isotype.
13. The fragment according to claim 8, characterized in that it comprises a V_H segment having an amino acid sequence defined by amino acid residues 1-122 of SEQ ID NO. 1 and/or a V_L segment having an amino acid sequence defined by amino acid residues 1-108 of SEQ ID NO. 2.
14. The fragment according to claim 13, characterized in that it comprises a Fv, single chain Fv, Fab or F-(ab')₂ fragment.
15. An isolated nucleic acid characterized in that it encodes a human monoclonal antibody or fragment according to any one of claims 1-14.
16. An isolated nucleic acid characterized in that it comprises:
 - a nucleotide sequence defined by base numbers 58-423 of SEQ ID NO. 1 or by a CDR encoding somatic variant thereof, and/or
 - 35 a nucleotide sequence defined by base numbers 67-390 of SEQ ID NO. 2 or by a CDR encoding somatic variant thereof; or
 - a functional equivalent of one or both of said nucleotide sequences.
17. An isolated nucleic acid according to claim 16, characterized in that it comprises a nucleotide sequence defined by base numbers 58-423 of SEQ ID NO. 1 and/or base numbers 67-390 of SEQ ID NO. 2.
18. A pharmaceutical composition characterized in that it comprises at least one human monoclonal antibody or fragment according to any one of claims 1-14 and a pharmaceutically acceptable carrier.
19. A pharmaceutical composition according to claim 18, characterized in that it comprises a human monoclonal antibody or fragment according to any one of claims 4 and 6-14.
20. The use of human monoclonal antibody or fragment according to any one of claims 4 and 6-14 in the manufacture of a medicament to treat inflammation.
21. A method for screening a solution for a desired human monoclonal antibody against a human protein characterized in that it comprises
 - (1) contacting the solution with labeled protein and polyclonal or monoclonal anti-human Ig coupled to a substrate or with labeled protein and protein G coupled to a substrate; and
 - 55 (2) determining if a desired human monoclonal antibody is present in the solution by detecting labeled protein in any immunoprecipitated product.

22. A method according to claim 21, characterized in that the solution is a collection of supernatants from a human B cell mixture.
23. A method according to any one of claims 21 or 22, characterized in that it further comprises the steps of
 - serially diluting a human B cell mixture giving a positive result to provide a purified mixture of human B cells or single B cell clones;
 - culturing said purified mixture of human B cells or single human B cell clones; and
 - screening supernatants from said cultured purified mixture of human B cells or single B cell clones by the method of claim 21 or 22 to determine if the desired human monoclonal antibody is present in the supernatants of said cultured purified mixture of human B cells or single B cell clones.
24. A method of producing a cDNA library enriched in DNA encoding a V_H and/or V_L chain of a human monoclonal antibody against a desired antigen characterized in that it comprises the steps of:
 - producing a CD40-crosslinked and EBV-transformed B cell population containing immortalized and/or activated B cells expressing said human monoclonal antibody;
 - cloning subpopulations of said immortalized and/or activated B cell population and identifying a subpopulation which contains immortalized and/or activated B cells expressing said human monoclonal antibody;
 - preparing a cDNA library using the mRNA from said subpopulation to create a repertoire of DNAs encoding at least the V_H and/or V_L chain of the human monoclonal antibodies expressed by said subpopulation of immortalized and/or activated B cells.
25. A method according to claim 24, characterized in that it further comprises:
 - identifying DNA within said library that encodes at least the V_H and/or V_L chain of the desired human monoclonal antibody; and
 - using said DNA to produce a human monoclonal antibody against the desired antigen or an antigen-binding fragment of such an antibody.
26. A method according to claim 24 or 25, characterized in that a population or subpopulation which contains immortalized and/or activated B cells expressing said human monoclonal antibody is identified by
 - (1) contacting supernatant from said population or subpopulation with labeled antigen and polyclonal or monoclonal anti-human Ig coupled to a substrate or with labeled antigen and protein G coupled to a substrate; and
 - (2) determining if a desired human monoclonal antibody is present in the supernatant by detecting labeled antigen in any immunoprecipitated product.
27. A method according to claim 24, 25 or 26, characterized in that the repertoire of DNAs are included in vectors capable of displaying said V_H and/or V_L chain on the surface of a host cell, host cells are transformed with said vectors, and host cells that display a V_H and/or V_L chain that binds to the desired antigen are identified by affinity binding to the desired antigen.
28. A method according to claim 24, 25 or 26, characterized in that DNAs encoding said V_H and V_L chains that bind to the desired antigen are operatively linked to DNA encoding any necessary constant region chains for a human immunoglobulin so as to create a DNA sequence encoding a heavy chain of a human monoclonal antibody against the desired antigen and a DNA sequence encoding a light chain of a human monoclonal antibody against the desired antigen.
29. A human B cell line established by EBV-transformation and CD40-crosslinking, characterized in that the established cell line produces a human monoclonal antibody according to any one of claims 1-12.
30. A human monoclonal antibody-producing clone of an established human B cell line according to claim 29.
31. A process for making a human monoclonal antibody against a human cytokine characterized in that it comprises the steps of
 - establishing an immortalized and/or activated human B cell population from a patient having

antibodies that bind to the human cytokine, said immortalization and/or activation comprising infecting the B cells with Epstein-Barr virus and crosslinking the CD40 of such B cells;

culturing said immortalized and/or activated B cells;

5 isolating multiple clones from such immortalized and/or activated B cells, each of which clones secretes a human monoclonal antibody that binds to the cytokine; and

using one or more of such clones to produce one or more human monoclonal antibody or a fragment thereof.

10 32. A process according to claim 31, characterized in that nucleic acid encoding the antibody or fragment is used to produce the desired antibody or fragment.

15 33. A process according to claim 31, characterized in that said clone is fused with a myeloma or heteromyeloma cell to produce a hybridoma that proliferates in culture and produces the desired antibody.

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FIGURE 1

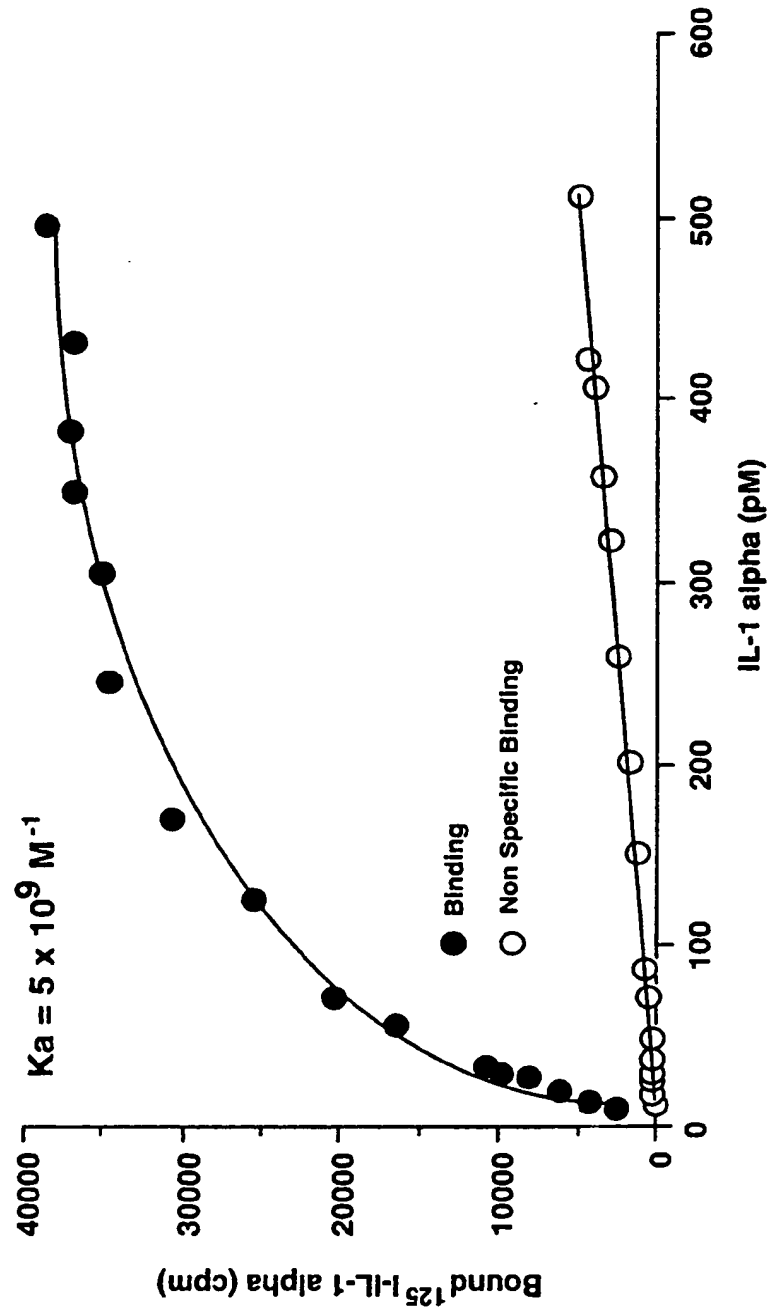


FIGURE 2

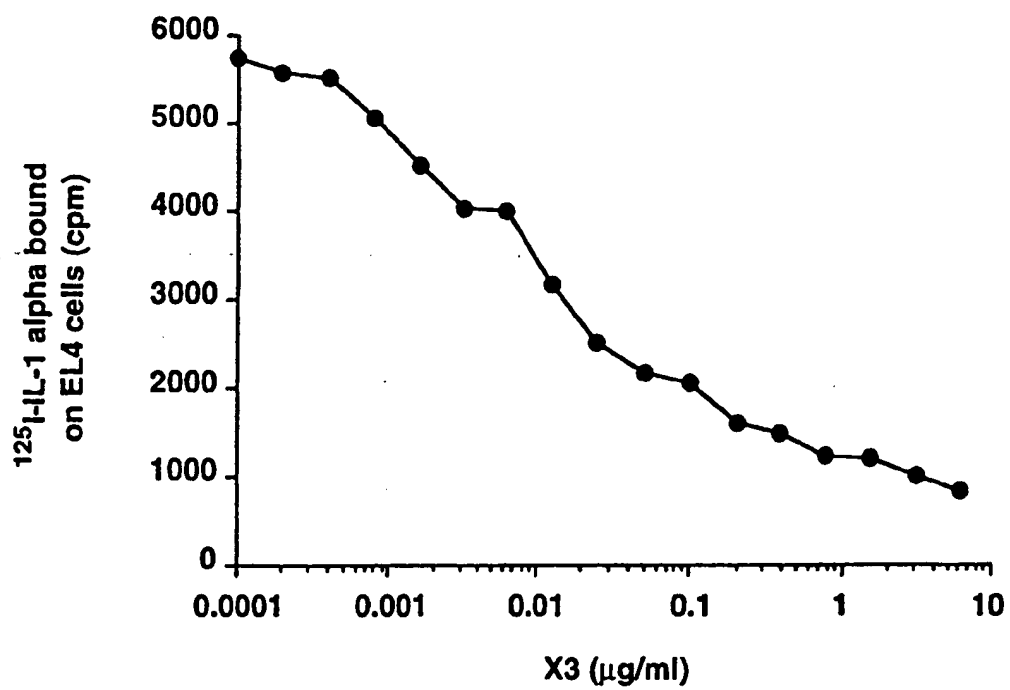
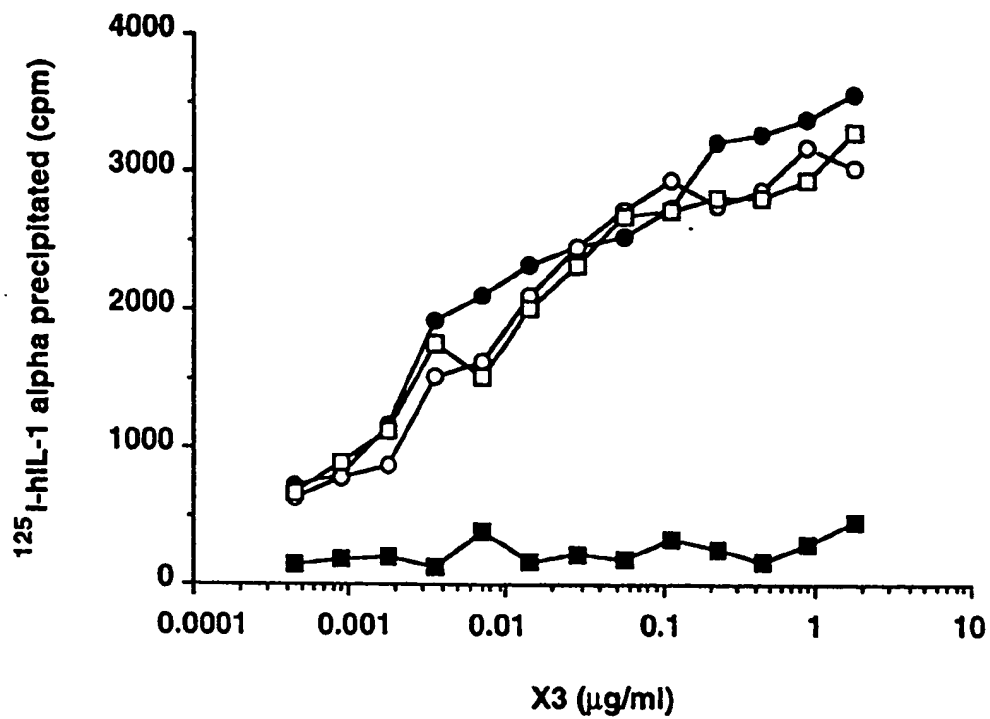


FIGURE 3**Protection:**

- none
- human rIL-1 alpha
- human rIL-1 beta
- human rIL-1 RA

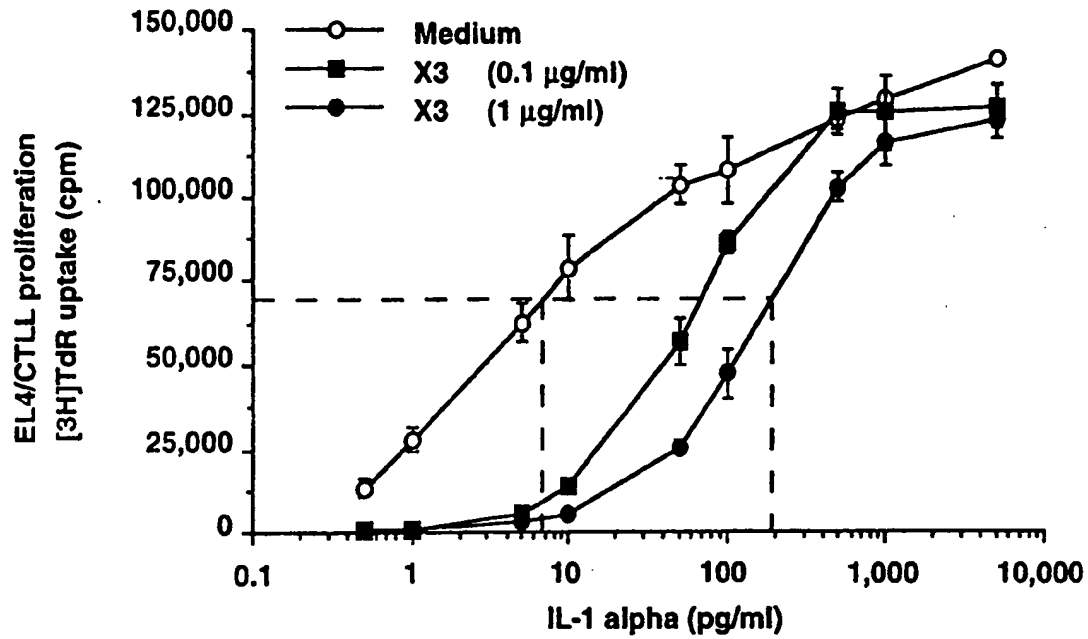
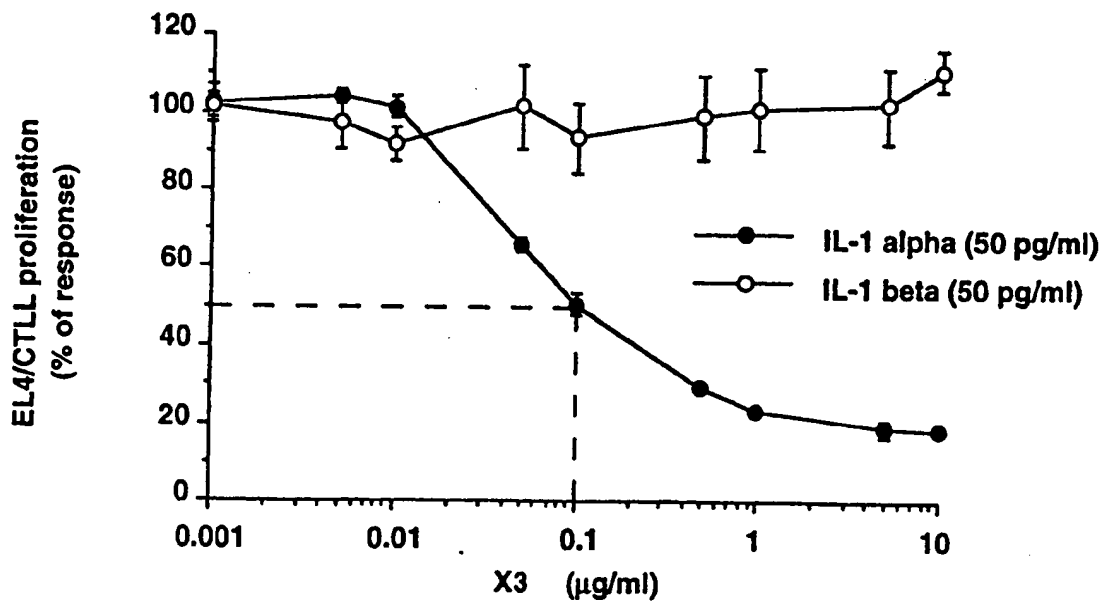
FIGURE 4A**FIGURE 4B**

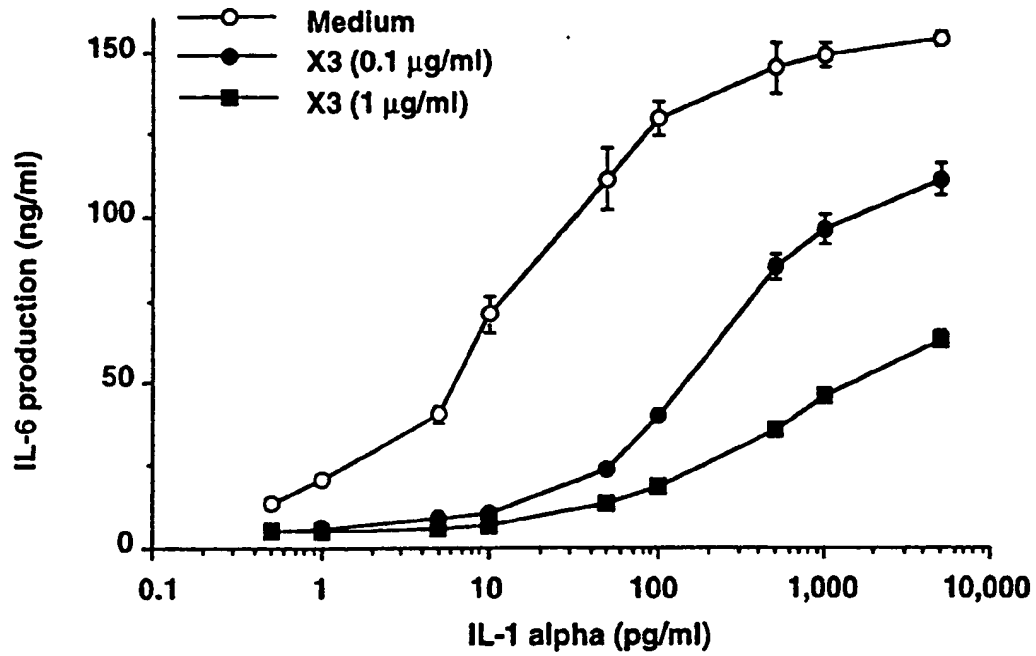
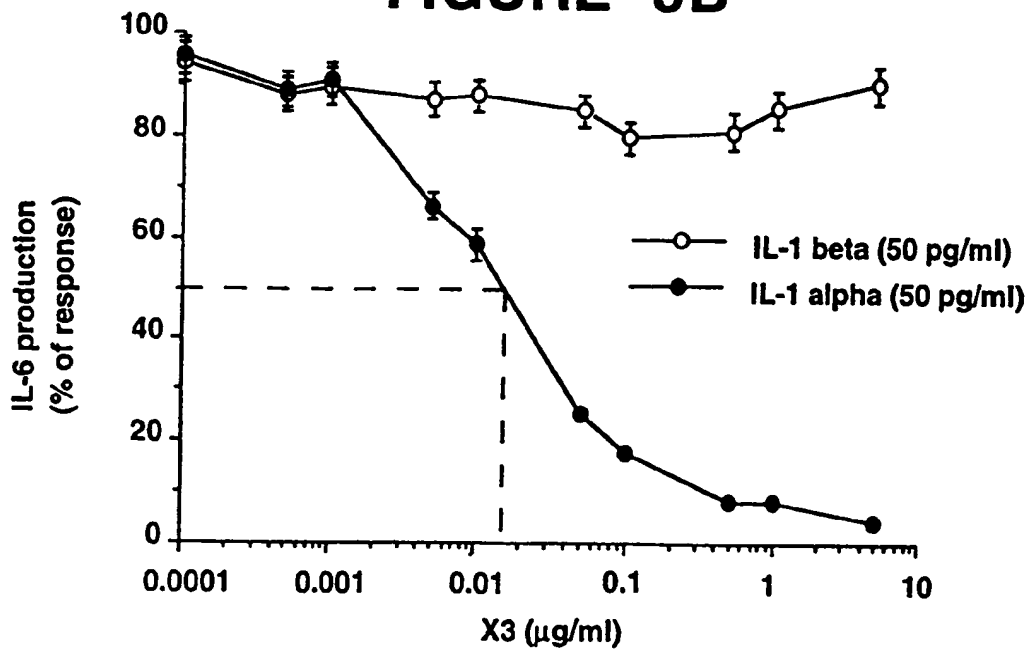
FIGURE 5A**FIGURE 5B**

FIGURE 6A

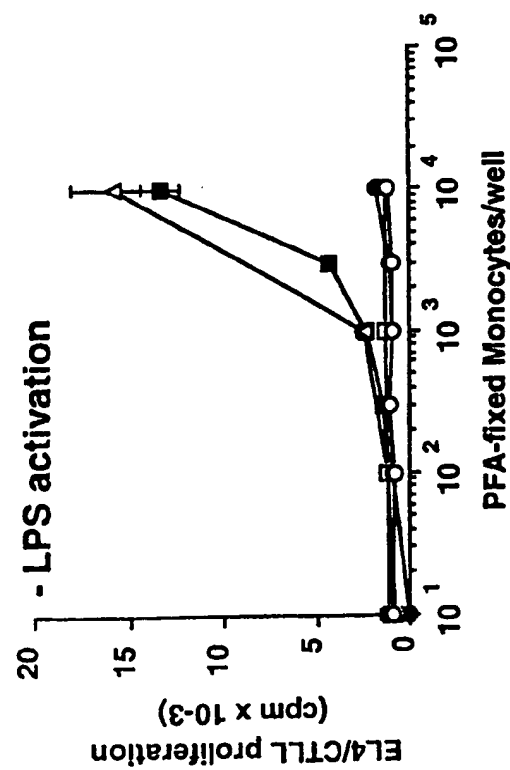


FIGURE 6B

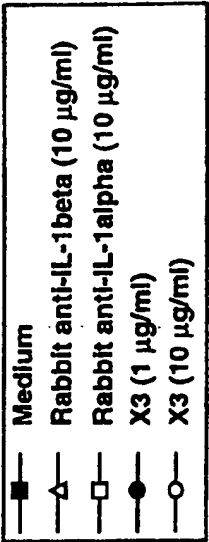
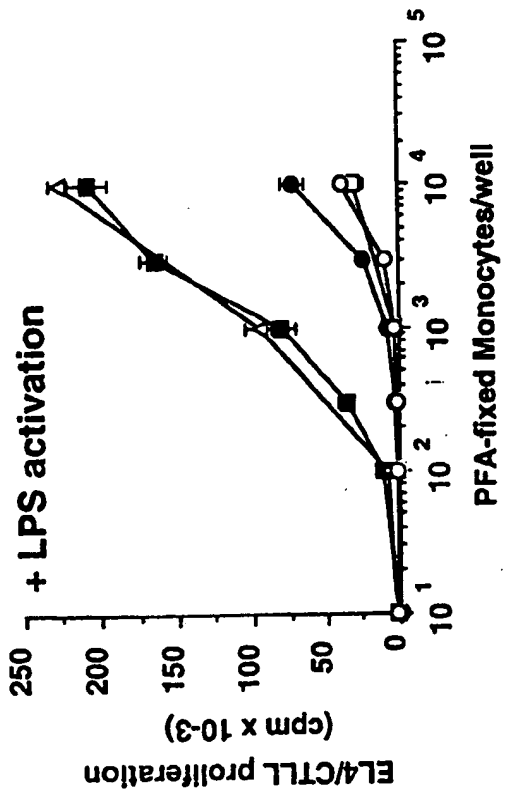


FIGURE 7

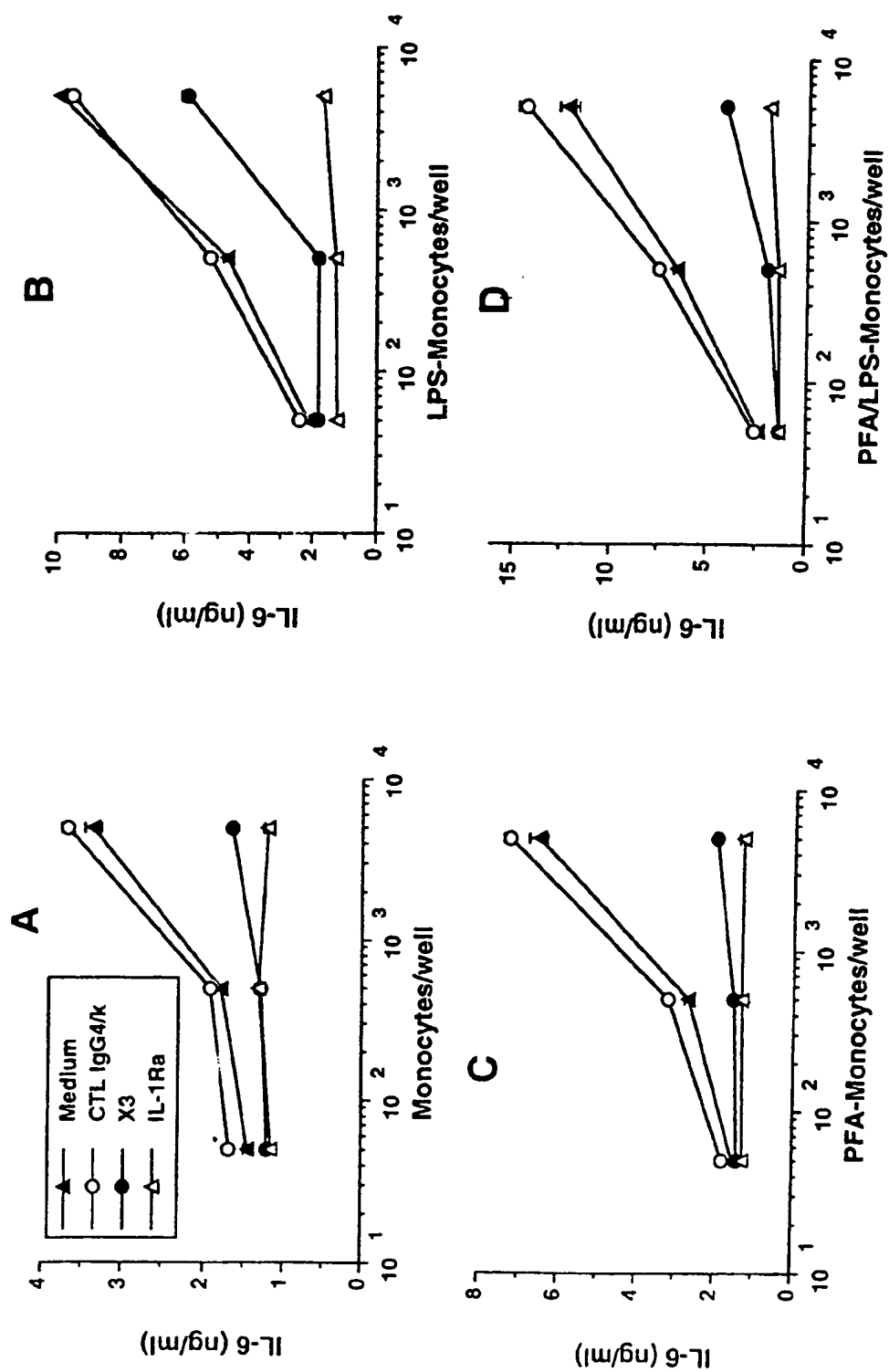


FIGURE 8A

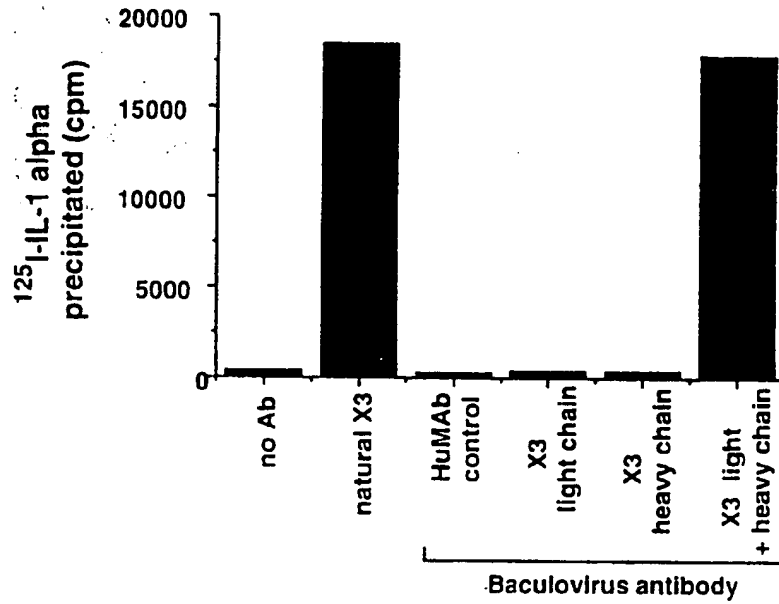
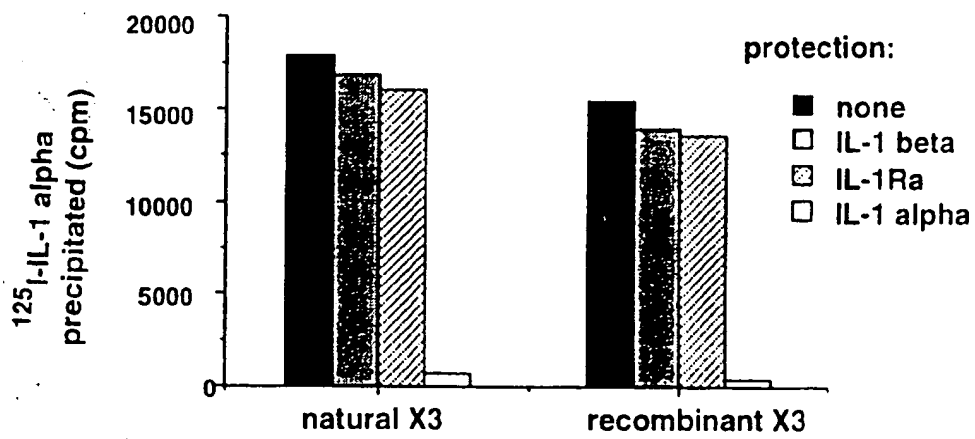


FIGURE 8B





European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 93 40 2846

| DOCUMENTS CONSIDERED TO BE RELEVANT | | | |
|---|---|---|--|
| Category | Citation of document with indication, where appropriate, of relevant passages | Relevant to claim | CLASSIFICATION OF THE APPLICATION (Int.Cl.6) |
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| Y | * example 2 * | 29-32 | G01N33/53 |
| A | --- | 5 | C07K14/52 |
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| Y | * page 3, line 5 - line 43 * | 29-32 | |
| A | * page 4, line 18 - line 31 * | 7-17 | |
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| A | * page 8, line 15 - line 30 * | 14 | |
| Y | EP-A-0 434 879 (LABORATOIRES UNICET) | 29-32 | |
| D | * claims 1-9 * & WO-A-91 09115 | | |
| X | EP-A-0 364 778 (OTSUKA PHARMACEUTICAL CO. LTD.) | 1-3 | |
| | * claims 1-9 * | | |
| | ----- | | |
| The present search report has been drawn up for all claims | | | |
| Place of search BERLIN | | Date of completion of the search 4 May 1995 | Examiner Gurdjian, D |
| CATEGORY OF CITED DOCUMENTS | | | |
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